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(54) Title: METHOD OF MAGNETIC RESONANCE INVESTIGATION

(57) Abstract

This invention provides a method of magnetic resonance investigation of a sample, preferably of a human or non-human animal body, said method comprising: (i) producing a hyperpolarised solution of a high T₁ agent by dissolving in a physiologically tolerable solvent a hyperpolarised solid sample of said high T₁ agent; (ii) where the hyperpolarisation of the solid sample of said high T₁ agent in step (i) is effected by means of a polarising agent, optionally separating the whole, substantially the whole, or a portion of said polarising agent from said high T₁ agent; (iii) administering said hyperpolarised solution to said sample; (iv) exposing said sample to a second radiation of a frequency selected to excite nuclear spin transitions in selected nuclei e.g. the MR imaging nuclei of the high T₁ agent; (v) detecting magnetic resonance signals from said sample; and (vi) optionally, generating an image, dynamic flow data, diffusion data, perfusion data, physiological data (e.g. pH, pO₂, pCO₂, temperature or ionic concentrations) or metabolic data from said detected signals, wherein said high T₁ agent in said hyperpolarised solution has a T₁ value (at a field strength in the range 0.01–5T and a temperature in the range 20–40 °C) of at least 5 seconds.

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METHOD OF MAGNETIC RESONANCE INVESTIGATION

This invention relates to a method of magnetic resonance imaging (MRI).

Magnetic resonance imaging (MRI) is a diagnostic technique that has become particularly attractive to physicians as it is non-invasive and does not involve exposing the patient under study to potentially harmful radiation such as X-rays.

In order to achieve effective contrast between MR images of the different tissue types in a subject, it has long been known to administer to the subject MR contrast agents (e.g. paramagnetic metal species) which effect relaxation times of the MR imaging nuclei in the zones in which they are administered or at which they aggregate. Contrast enhancement has also been achieved by utilising the "Overhauser effect" in which an esr transition in an administered paramagnetic species (hereinafter an OMRI contrast agent) is coupled to the nuclear spin system of the imaging nuclei. The Overhauser effect (also known as dynamic nuclear polarisation) can significantly increase the population difference between excited and ground nuclear spin states of selected nuclei and thereby amplify the MR signal intensity by a factor of a hundred or more allowing OMRI images to be generated rapidly and with relatively low primary magnetic fields. Most of the OMRI contrast agents disclosed to date are radicals which are used to effect polarisation of imaging nuclei *in vivo*.

Techniques are now being developed which involve *ex vivo* polarisation of agents containing MR imaging nuclei, prior to administration and MR signal measurement. Such techniques may involve the use of polarising agents, for example conventional OMRI contrast agents or hyperpolarised gases to achieve *ex*

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vivo polarisation of administrable MR imaging nuclei. By polarising agent is meant any agent suitable for performing ex vivo polarisation of an MR imaging agent.

The ex vivo method has inter alia the advantage that it is possible to avoid administering the whole of, or substantially the whole of, the polarising agent to the sample under investigation, whilst still achieving the desired polarisation. Thus the method is less constrained by physiological factors such as the constraints imposed by the administrability, biodegradability and toxicity of OMRI contrast agents in in vivo techniques.

It has now been found that ex vivo methods of magnetic resonance imaging may be improved by using polarised MR imaging agents comprising nuclei capable of emitting magnetic resonance signals in a uniform magnetic field (eg MR imaging nuclei such as ^{13}C or ^{15}N nuclei) and capable of exhibiting a long T_1 relaxation time, preferably additionally a long T_2 relaxation time. Such agents will be referred to hereinafter as "high T_1 agents". Typically the molecules of a high T_1 agent will contain MR imaging nuclei in an amount greater than the natural abundance of said nuclei in said molecules (i.e. the agent will be enriched with said nuclei).

Thus viewed from one aspect the present invention provides a method of magnetic resonance investigation of a sample, preferably of a human or non-human animal body (eg. a mammalian, reptilian or avian body), said method comprising:

- (i) producing a hyperpolarised solution of a high T_1 agent by dissolving in a physiologically tolerable solvent a hyperpolarised solid sample of said high T_1 agent;
- (ii) where the hyperpolarisation of the solid sample of said high T_1 agent in step (i) is effected by means of a polarising agent, optionally separating the whole, substantially the whole, or a portion of said polarising

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- agent from said high T_1 agent;
- (iii) administering said hyperpolarised solution to said sample;
 - (iv) exposing said sample to a second radiation of a frequency selected to excite nuclear spin transitions in selected nuclei eg the MR imaging nuclei of the high T_1 agent;
 - (v) detecting magnetic resonance signals from said sample; and
 - (vi) optionally, generating an image, dynamic flow data, diffusion data, perfusion data, physiological data (eg. pH, pO_2 , pCO_2 , temperature or ionic concentrations) or metabolic data from said detected signals,

wherein said high T_1 agent in said hyperpolarised solution has a T_1 value (at a field strength in the range 0.01-5T and a temperature in the range 20-40°C) of at least 5 seconds.

Thus the invention involves the sequential steps of producing a hyperpolarised solution from a hyperpolarised solid sample of a high T_1 agent comprising nuclei capable of exhibiting a long T_1 relaxation time, administration of the hyperpolarised solution of the high T_1 agent (preferably in the absence of a portion of, more preferably substantially the whole of, any polarising agent), and conventional *in vivo* MR signal generation and measurement. The MR signals obtained in this way may be conveniently converted by conventional manipulations into 2-, 3- or 4-dimensional data including flow, diffusion, physiological or metabolic data.

By "hyperpolarised" we mean polarised to a level over that found at room temperature and 1T, preferably polarised to a polarisation degree in excess of 0.1%, more preferably 1%, even more preferably 10%.

Polarization is given by the equation

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$$P = \left| \frac{N\alpha - N\beta}{N\alpha + N\beta} \right|$$

which at equilibrium is equal to

$$\frac{1 - \exp(-\gamma\hbar B_0/kT)}{1 + \exp(-\gamma\hbar B_0/kT)}$$

where $N\alpha$ is the number of spins in nuclear spin state α (e.g. $+\frac{1}{2}$);

$N\beta$ is the number of spins in nuclear spin state β (e.g. $-\frac{1}{2}$);

γ is the magnetogyric ratio for the isotopic nucleus in question, e.g. ^{13}C);

\hbar is Planck's constant divided by 2π ;

B_0 is the magnetic field;

k is Boltzmann's constant; and

T is temperature in kelvin.

Thus P has a maximum value of 1 (100% polarization) and a minimum value of 0 (0% polarization).

By "physiologically tolerable solvent" we mean any solvent, solvent mixture or solution that is tolerated by the human or non-human animal body, e.g. water, aqueous solutions such as saline, perfluorocarbons, etc.

One embodiment of the invention provides a method as described above wherein the hyperpolarised solid sample of said high T_1 agent retains its polarisation when transported in a magnetic field and at low temperature; in this way the agent can be hyperpolarised at a site remote from its end use and transported to its place of use in a magnetic field and at a low temperature and there dissolved and administered.

In the embodiment referred to above, the magnetic field is preferably greater than 10 mT, more preferably greater than 0.1T, even more preferably greater than 0.5T, yet more preferably greater than 1T. By "low

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"temperature" we preferably mean lower than 80K, more preferably lower than 4.2K, most preferably lower than 1K.

A further embodiment of the invention provides a method as described above wherein the hyperpolarised solution thus formed retains its polarisation when transported in a magnetic field. In this latest embodiment, the magnetic field is preferably greater than 10 mT, more preferably greater than 0.1T, even more preferably greater than 0.5T, yet more preferably greater than 1T.

A yet further embodiment of the invention provides a method as described above wherein a magnetic field is present during the dissolution stage. In this latest embodiment, the magnetic field is preferably greater than 10 mT, more preferably greater than 0.1T, even more preferably greater than 0.5T, yet more preferably greater than 1T.

Suitable high T_1 agents may contain nuclei such as protons. However other non-zero nuclear spin nuclei may be useful (eg ^{19}F , ^3Li , ^{13}C , ^{15}N , ^{29}Si or ^{31}P , as well as ^1H), preferably ^1H , ^{13}C , ^{15}N , ^{19}F , ^{29}Si and ^{31}P nuclei, with ^{13}C and ^{15}N nuclei being particularly preferred. In this event the MR signals from which the image is generated will be substantially only from the high T_1 agent itself. Nonetheless, where the polarised high T_1 agent is present in high concentration in administrable media, there may be significant enough transfer of magnetisation to the protons to be able to perform ^1H -MRI on the protons of the media. Similarly, the polarised high T_1 agent may have a significant enough effect on *in vivo* protons for conventional ^1H -MRI to be carried out on those protons.

Where the MR imaging nuclei is other than a proton (eg ^{13}C or ^{15}N), there will be essentially no interference from background signals (the natural abundance of ^{13}C and ^{15}N being negligible) and image contrast will be advantageously high. This is especially true where the

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high T_1 agent itself is enriched above natural abundance. Thus the method according to the invention has the benefit of being able to provide significant spatial weighting to a generated image. In effect, the administration of a polarised high T_1 agent to a selected region of a sample (eg by injection) means that the contrast effect may be localised to that region. The precise effect of course depends on the extent of biodistribution over the period in which the high T_1 agent remains significantly polarised. In general, specific body volumes (i.e. regions of interest such as the vascular system or specific organs such as the brain, kidney, heart or liver) into which the agent is administered may be defined with improved signal to noise (particularly improved contrast to noise) properties of the resulting images in these volumes.

In one embodiment, a "native image" of the sample (e.g. body) (ie. one obtained prior to administration of the high T_1 agent or one obtained for the administered high T_1 agent without prior polarisation as in a conventional MR experiment) may be generated to provide structural (eg. anatomical) information upon which the image obtained in the method according to the invention may be superimposed. A "native image" is generally not available where ^{13}C or ^{15}N is the imaging nucleus because of the low abundance of ^{13}C and ^{15}N in the body. In this case, a proton MR image may be taken to provide the anatomical information upon which the ^{13}C or ^{15}N image may be superimposed.

The high T_1 agent should of course be physiologically tolerable or be capable of being provided in a physiologically tolerable, administrable form. Preferred high T_1 agents are soluble in aqueous media (eg. water) and are of course non-toxic where the intended end use is *in vivo*.

Conveniently, the high T_1 agent once polarised will remain so for a period sufficiently long to allow the

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imaging procedure to be carried out in a comfortable time span. Generally sufficient polarisation will be retained by the high T₁ agent in its administrable form (eg. in injection solution) if it has a T₁ value (at a field strength of 0.01-5T and a temperature in the range 20-40°C) of at least 5s, more preferably at least 10s, especially preferably 30s or longer, more especially preferably 70s or more, yet more especially preferably 100s or more (for example at 37°C in water at 1T and a concentration of at least 1mM). The high T₁ agent may be advantageously an agent with a long T₂ relaxation time.

The long T₁ relaxation time of certain ¹³C nuclei is particularly advantageous and certain high T₁ agents containing ¹³C nuclei are therefore preferred for use in the present method. The γ-factor of carbon is about ¼ of the γ-factor for hydrogen resulting in a Larmor frequency of about 10 MHz at 1 T. The rf-absorption and reflections in a patient is consequently and advantageously less than in water (proton) imaging. The signal-to-noise ratio is found to be independent of the MRI field strength when the corresponding frequency is higher than a few MHz. Preferably the polarised high T₁ agent has an effective ¹³C nuclear polarisation corresponding to the one obtained at thermal equilibrium at 300K in a field of 0.1T or more, more preferably 25T or more, particularly preferably 100T or more, especially preferably 5000T or more (for example 50 kT).

When the electron cloud of a given molecule interacts with atoms in surrounding tissue, the shielding of the atom responsible for the the MR signal is changed giving rise to a shift in the MR frequency ("the chemical shift effect"). When the molecule is metabolised, the chemical shift will be changed and high T₁ agents in different chemical surroundings may be visualised separately using pulses sensitive to chemical shift. When the frequency difference between high T₁ molecules in different surroundings is 10 Hz or higher,

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preferably 20 Hz or higher, most preferably 150 Hz or higher (corresponding to 3.5ppm or higher at 1T), the two components may be excited separately and visualised in two images. Standard chemical shift selective excitation pulses may then be utilised. When the frequency separation is less, the two components may not be separated by using frequency selective rf-pulses. The phase difference created during the time delay after the excitation pulse and before the detection of the MR signal may then be used to separate the two components. Phase sensitive imaging pulse sequence methods (Dixon, Radiology, 1984, 153: 189-194 and Sepponen, Mag Res. Imaging, 3, 163-167, 1985) may be used to generate images visualising different chemical surroundings or different metabolites. The long T₂ relaxation time which may be a characteristic of a high T₁ agent will under these circumstances make it possible to use long echo times (TE) and still get a high signal-to-noise ratio. Thus an important advantage of the high T₁ agents used in the present method is that they exhibit a chemical shift dependent on the local composition of the body in which they are localised. Preferred high T₁ agents will exhibit at 1T a chemical shift of more than 2ppm, preferably more than 10ppm depending on whether the high T₁ agent is localised inside or outside the vascular system. More preferred high T₁ agents will exhibit a chemical shift of more than 2 ppm, preferably more than 10 ppm, per 2 pH units or per Kelvin or upon being metabolised. High T₁ agents containing polarised ¹³C nuclei (or ¹⁵N nuclei) exhibit large changes in chemical shift in response to physiological changes (eg. pH, pO₂, pCO₂, redox potential, temperature or ionic concentrations of for example Na⁺, K⁺, Ca²⁺) or metabolic activity and therefore may be used to monitor these parameters.

Solid high T₁ agents (e.g. ¹³C or ¹⁵N enriched solids) may exhibit very long T₁ relaxation times and for

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this reason are especially preferred for use in the present method. The T_1 relaxation time may be several hours in the bulk phase, although this may be reduced by reduction of grain size and/or addition of paramagnetic impurities eg. molecular oxygen. The long relaxation time of solids advantageously allows the procedure to be conveniently carried out with less haste and is particularly advantageous in allowing the polarised solid high T_1 agent to be stored or transported prior to pharmaceutical formulation and administration. In one embodiment, the polarised high T_1 agent may be stored at low temperature and prior to administration, the high T_1 agent may be rapidly warmed to physiological temperatures using conventional techniques such as infrared or microwave radiation or simply by adding hot, sterile administrable media eg saline.

For *in vivo* use, a polarised solid high T_1 agent is dissolved in administrable media (eg water or saline), administered to a subject and conventional MR imaging performed. Thus solid high T_1 agents are preferably rapidly soluble (eg. water soluble) to assist in formulating administrable media. Preferably the high T_1 agent should dissolve in a physiologically tolerable carrier (eg water or Ringers solution) to a concentration of at least 1mM at a rate of 1mM/3 T_1 or more, particularly preferably 1mM/2 T_1 or more, especially preferably 1mM/ T_1 or more. Where the solid high T_1 agent is frozen, the administrable medium may be heated, preferably to an extent such that the temperature of the medium after mixing is close to 37°C.

A polarised high T_1 agent may be administered (either alone or with additional components such as additional high T_1 agents) in liquid form. The retention of polarisation in a liquid medium vis-a-vis a gas medium is significantly greater. Thus while T_1 and T_2 are in general shorter for the liquid, the T_2^* effect due to diffusion is 10^5 times less significant for the

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liquid. Consequently for gaseous high T_1 agents the imaging sequence used generally has to be FLASH or GRASS while in contrast, more efficient imaging sequences may be used for liquids. For example, liquids generally have slower diffusion which makes it possible to use sequences such as echo planar imaging (EPI). The overall technique will be faster and yield better resolution (voxel size < 1mm) than conventional techniques (voxel size approx. 1-5mm) at current acquisition times. It will give good images at all fields including in low field (eg. 0.01-0.5T) machines.

Unless the hyperpolarised agent is stored (and/or transported) at low temperature and in an applied field as described above, since the method of the invention should be carried out within the time that the hyperpolarised solution of the high T_1 agent remains significantly polarised, it is desirable for administration of the polarised high T_1 agent to be effected rapidly and for the MR measurement to follow shortly thereafter. The preferred administration route for the polarised high T_1 agent is parenteral eg by bolus injection, by intravenous, intraarterial or peroral injection. The injection time should be equivalent to $5T_1$ or less, preferably $3T_1$ or less, particularly preferably T_1 or less, especially $0.1T_1$ or less. The lungs may be imaged by spray, eg by aerosol spray.

The high T_1 agent should be preferably enriched with nuclei (eg. ^{15}N and/or ^{13}C nuclei) having a long T_1 relaxation time. Preferred are ^{13}C enriched high T_1 agents having ^{13}C at one particular position (or more than one particular position) in an amount in excess of the natural abundance, i.e. above about 1%. Preferably such a single carbon position will have 5% or more ^{13}C , particularly preferably 10% or more, especially preferably 25% or more, more especially preferably 50% or more, even more preferably in excess of 99% (e.g. 99.9%). The ^{13}C nuclei should preferably amount to >2%

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of all carbon atoms in the compound. The high T₁ agent is preferably ¹³C enriched at one or more carbonyl or quaternary carbon positions, given that a ¹³C nucleus in a carbonyl group or in certain quaternary carbons may have a T₁ relaxation time typically of more than 2s, preferably more than 5s, especially preferably more than 30s. Preferably the ¹³C enriched compound should be deuterium labelled, especially adjacent the ¹³C nucleus.

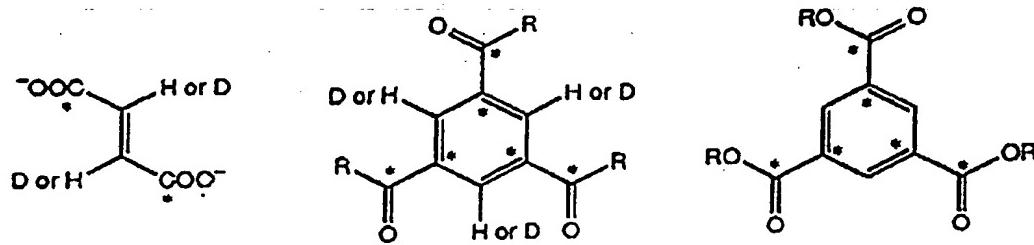
Viewed from a further aspect the present invention provides a composition comprising a hyperpolarised solution of a polarised ¹³C, ¹⁵N, ¹⁹F, ²⁹Si, ³¹P or ¹H enriched compound together with one or more physiologically acceptable carriers or excipients.

Viewed from a further aspect the present invention provides a contrast medium comprising a hyperpolarised solution of a polarised high T₁ agent being enriched with ¹³C, ¹⁵N, ¹⁹F, ²⁹Si, ³¹P or ¹H nuclei having a T₁ relaxation time of 2s or more, preferably 10 secs or more, more preferably 30 secs or more, especially preferably 60 secs or more in solution at magnetic fields of 0.005-10T, preferably 0.01-10T, together with one or more physiologically acceptable carriers or excipients.

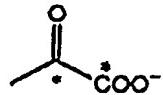
Preferred ¹³C enriched compounds are those in which the ¹³C nucleus is surrounded by one or more non-MR active nuclei such as O, S, C or a double bond. Specifically preferred ¹³C enriched agents are ¹³CO₃²⁻ and H¹³CO₃⁻ (sodium salt for injection and calcium or potassium salt for polarisation).

Also preferred are the following types of compound (* denotes ¹³C enriched positions):

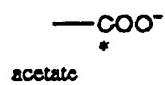
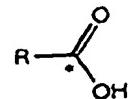
(1) carboxyl compounds comprising 1 to 4 carboxyl groups:



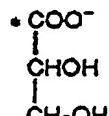
- 12 -



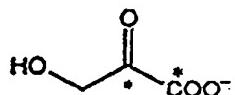
(including deuterated and
 ^{18}F labelled analogues)



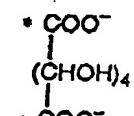
glyoxylic acid



glyceric acid



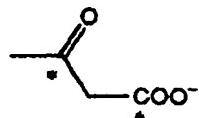
hydroxypyruvic acid



gluconic acid



oxalic acid

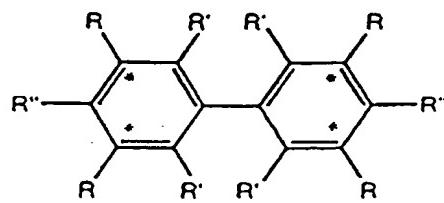
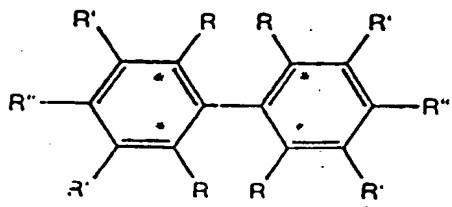
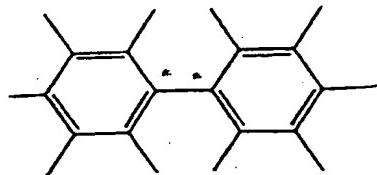
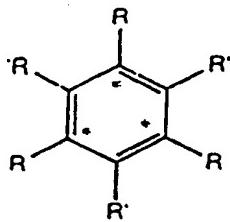


acetoacetate

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(wherein R represents any straight or branched chain hydrocarbon moiety, preferably a highly substituted carbon atom, especially preferably a quaternary carbon) and esters, isomers, especially stereoisomers and rotamers, thereof;

(2) substituted mono and biaryl compounds:



(wherein each group R or R' is independently a hydrogen atom, an iodine atom, a ¹⁹F atom or a hydrophilic moiety M being any of the non-ionizing groups conventionally used to enhance water solubility within the field of triiodophenyl X-ray contrast agents including for example a straight chain or branched C₁₋₁₀-alkyl group, preferably a C₁₋₅ group, optionally with one or more CH₂ or CH moieties replaced by oxygen or nitrogen atoms and optionally substituted by one or more groups selected from oxo, hydroxy, amino, carboxyl derivative, and oxo substituted sulphur and phosphorus atoms).

Particular examples of group M include polyhydroxyalkyl, hydroxyalkoxyalkyl and hydroxypolyalkoxyalkyl and such groups attached to the phenyl group via an amide linkage such as

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hydroxyalkylaminocarbonyl, N-alkyl-hydroxyalkylaminocarbonyl and bis-hydroxyalkylaminocarbonyl groups. Preferred among such M groups are those containing 1, 2, 3, 4, 5 or 6, especially 1, 2 or 3, hydroxy groups, e.g.

- CONH-CH₂CH₂OH
- CONH-CH₂CHOHCH₂OH
- CONH-CH(CH₂OH)₂
- CON(CH₂CH₂OH)₂

as well as other groups such as

- CONH₂
- CONHCH₃
- OCOCH₃
- N(COCH₃)H
- N(COCH₃)C₁₋₃-alkyl
- N(COCH₃)-mono, bis or tris-hydroxy C₁₋₄-alkyl
- N(COCH₂OH)-mono, bis or tris-hydroxy C₁₋₄-alkyl
- N(COCH₂OH)₂
- CON(CH₂CHOHCH₂OH)(CH₂CH₂OH)
- CONH-C(CH₂OH)₃ and
- CONH-CH(CH₂OH)(CHOHCH₂OH).

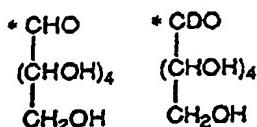
In general, the M groups will preferably each comprise a polyhydroxy C₁₋₄-alkyl group, such as C₁₋₄-alkyl groups substituted by 1, 2, 3 or 4 hydroxy groups (e.g. hydroxymethyl, 2-hydroxyethyl, 2,3-bishydroxy-propyl, 1,3-bishydroxyprop-2-yl, 2,3,4-trihydroxybutyl, and 1,2,4-trihydroxybut-2-yl) optionally connected to the phenyl ring via a CO, SO or SO₂ group (e.g. COCH₂OH or SO₂CH₂OH).

Preferred compounds are those in which two or three non-adjacent R groups in the or each C₆R₅ moiety are iodine and at least one, and preferably two or three, R groups in the or each C₆R₅ moiety are M or M₁ moieties; each M independently is a non-ionic hydrophilic moiety; and each M₁ independently represents a C₁₋₄-alkyl group.

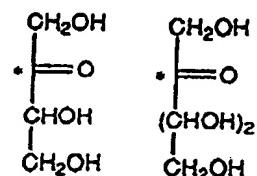
- 15 -

substituted by at least one hydroxyl group and optionally linked to the phenyl ring via a carbonyl, sulphone or sulphoxide group, at least one R group, preferably at least two R groups and especially preferably at least one R group in the or each C₆R₅ moiety, being an M₁ moiety. Especially preferred are the compounds disclosed in WO-A-96/09282.

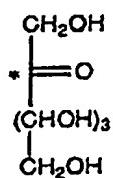
(3) sugars:



glucose

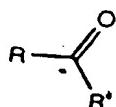


erythrose xylulose



fructose

(4) ketones:



(wherein R and R' are as defined above)

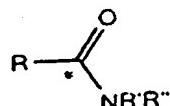
- 16 -

(5) ureas:

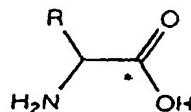


urea

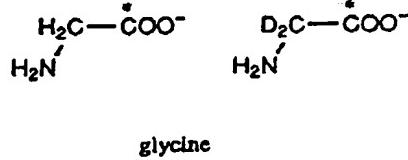
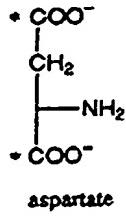
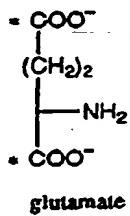
(6) amides:



(7) amino acids:

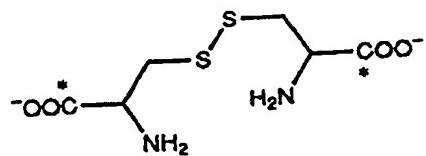


and peptides and proteins labelled in the carbonyl position, particularly those known in the art to be useful for targetting tumour cells. Of the proteins, albumin is especially preferred. Polymers are also useful, particularly those with low toxicity (eg polylysine) and those with many carboxyl groups (eg polyglutamic acid). The following amino acids are especially preferred:



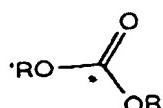
glycine

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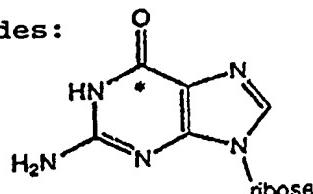


cysteine

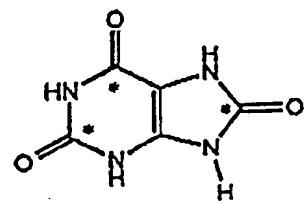
(8) carbonates:



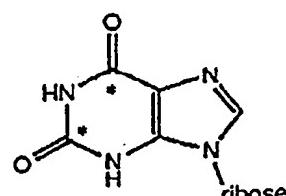
(9) nucleotides:



guanylic acid

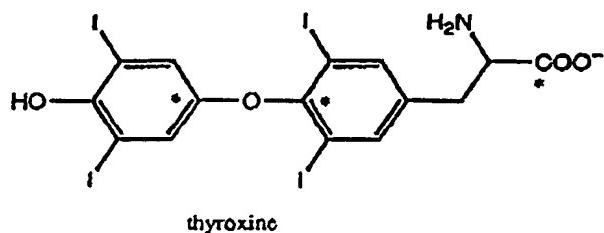


uric acid



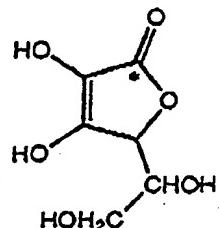
xanthyllic acid

(10) tracers:

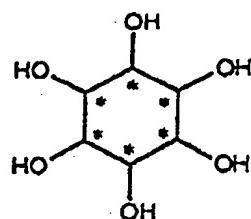


thyroxine

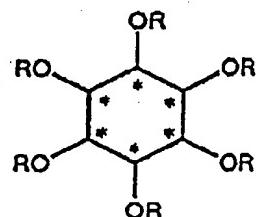
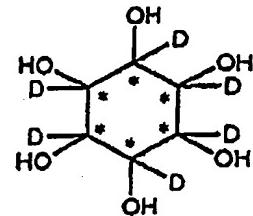
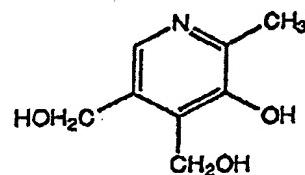
- 18 -



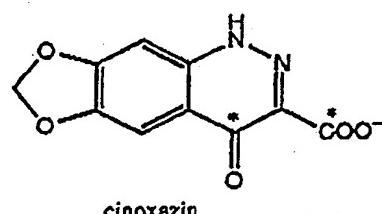
ascorbic acid



inositol

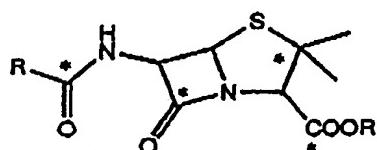
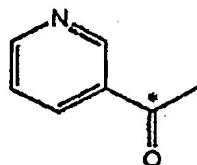
inositolhexanicotinate
(hexanicil, Astra)

pyridoxine

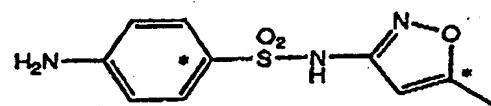


cinoxazin

R =

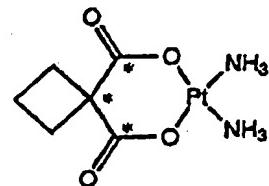


penicillin derivatives

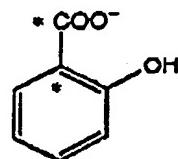


Sulfonamide

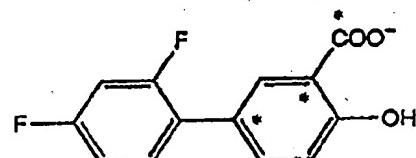
- 19 -



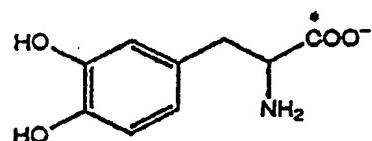
Carboplatinum



salicylate



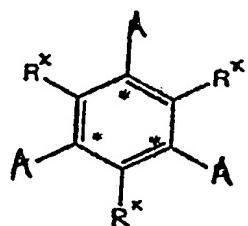
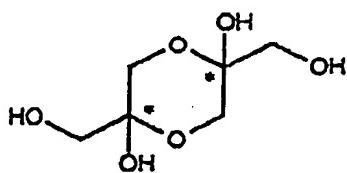
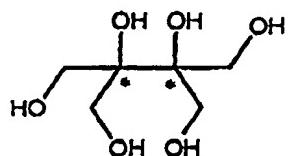
diflunisal



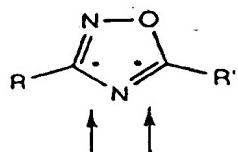
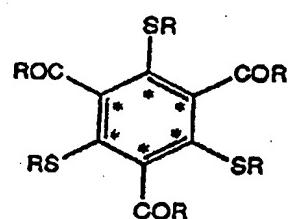
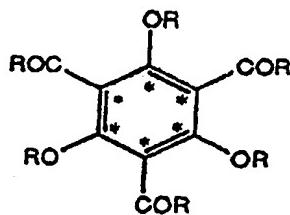
DOPA

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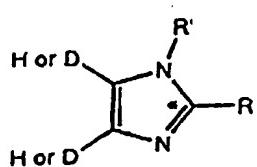
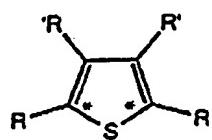
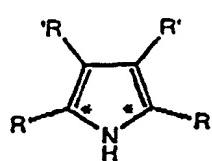
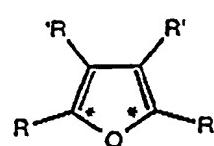
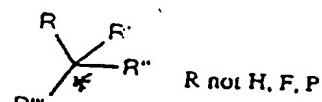
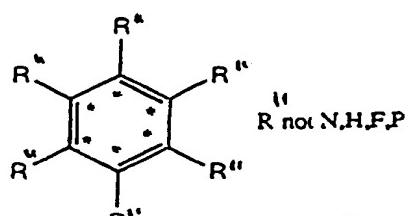
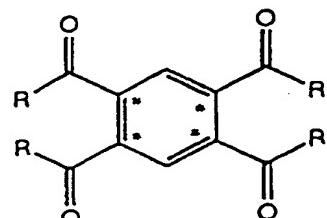
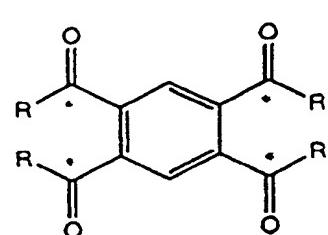
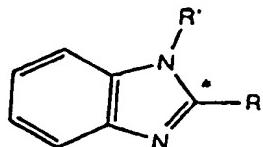
and (11) compounds such as:



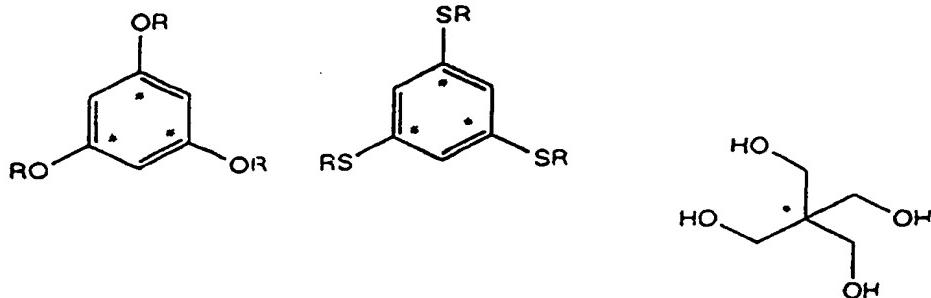
(wherein R denotes any of the conventional side chains suitable for use in X-ray contrast agents and A denotes I, D, OR, RC=O or ^{19}F)



(choose one of the two ^{13}C)



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In any of the above definitions, unless otherwise specified R, R', R'' and R''' denote any suitable substituent, preferably a substituent bound by a non-magnetic nucleus.

The partly or wholly deuterated or ¹⁹F analogues of any of these compounds are particularly preferred.

Certain of the above-mentioned ¹³C enriched compounds are novel *per se* and form a further aspect of the invention. Compounds which are water soluble are particularly preferred.

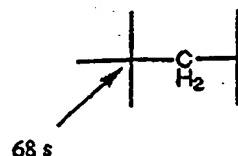
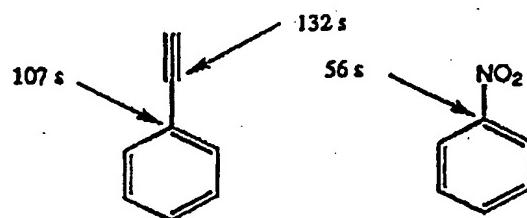
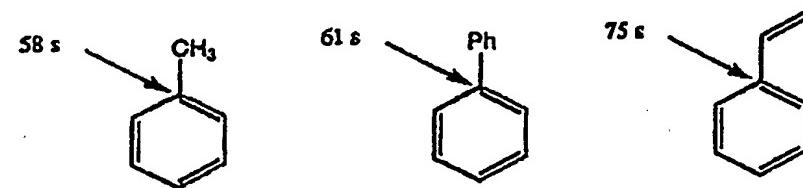
In general, ¹³C enriched amino acids and any known contrast agents from the fields of X-ray contrast agents and MRI contrast agents (the chelating agent without the metal counterion eg conventional Gd chelating agents without Gd) are preferred as high T₁ agents.

Intermediates in normal metabolic cycles such as the citric acid cycle eg. fumaric acid and pyruvic acid are preferred for the imaging of metabolic activity.

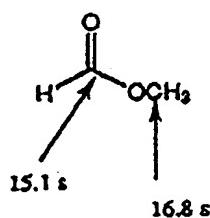
T₁ values for ¹³C enriched compounds useful in the invention are reported in the literature or may be routinely determined. Examples include:

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(a) non-water soluble (i.e soluble in an organic solvent)

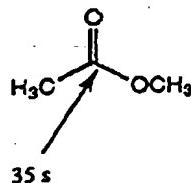


68 s



15.1 s

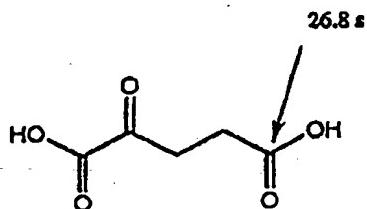
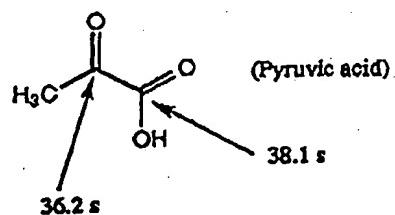
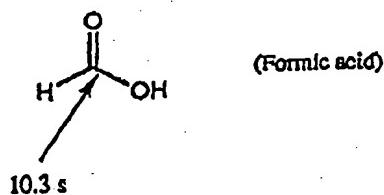
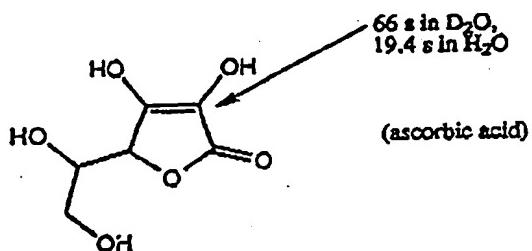
16.8 s

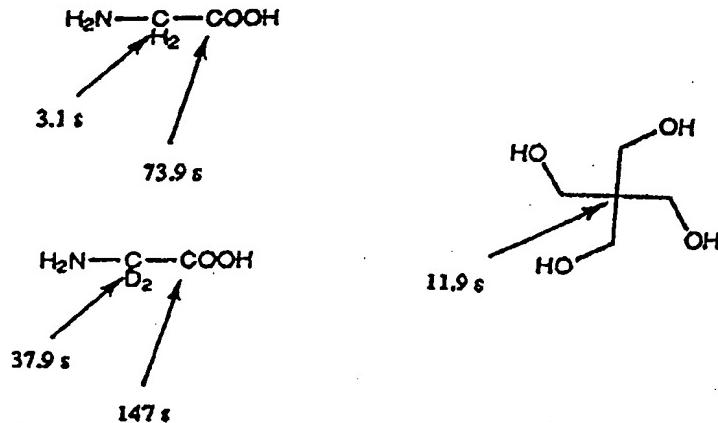


35 s

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(b) water soluble





Hyperpolarisation may be carried out by any known method and by way of example three such methods are described hereinbelow. It is envisaged that, in the method according to the invention, the level of polarisation achieved should be sufficient to allow the hyperpolarised solution of the high T₁ agent to achieve a diagnostically effective contrast enhancement in the sample to which it is subsequently administered in whatever form. In general, it is desirable to achieve a level of polarisation which is at least a factor of 2 or more above the field in which MRI is performed, preferably a factor of 10 or more, particularly preferably 100 or more and especially preferably 1000 or more, eg. 50000.

In a first embodiment of the method according to the invention, hyperpolarisation of the MR imaging nuclei is effected by an OMRI contrast agent. In this embodiment, step (i) of the method comprises:

- (a) bringing an OMRI contrast agent and a high T₁ agent into contact in a uniform magnetic field (the primary magnetic field B_0);
- (b) exposing said OMRI contrast agent to a first radiation of a frequency selected to excite electron

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spin transitions in said OMRI contrast agent; and

(c) dissolving in a physiologically tolerable solvent said high T₁ agent.

It is preferred that the OMRI contrast agent and high T₁ agent are present as a composition during polarisation.

Dynamic nuclear polarisation may be attained by three possible mechanisms: (1) the Overhauser effect, (2) the solid effect and (3) thermal mixing effect (see A.Abragam and M. Goldman, Nuclear Magnetism: order and disorder, Oxford University Press, 1982). The Overhauser effect is a relaxation driven process that occurs when the electron-nucleus interaction is time-dependent (due to thermal motion or relaxation effects) on the time scale of the inverse electron Larmor frequency or shorter. Electron-nuclear cross-relaxation results in an exchange of energy with the lattice giving rise to an enhanced nuclear polarisation. The overall enhancement depends on the relative strength of the scalar and dipolar electron-nuclear interaction and the microwave power. For static systems both thermal mixing and the solid effect are operative. In the solid effect, the electron spin system is irradiated at a frequency that corresponds to the sum or difference of the electronic and nuclear Larmor frequencies. The nuclear Zeeman reservoir absorbs or emits the energy difference and its spin temperature is modified, resulting in an enhanced nuclear polarisation. The efficiency depends on the transition probabilities of otherwise forbidden transitions that are allowed due to the mixing of nuclear states by non-secular terms of the electron-nuclear dipolar interaction. Thermal mixing arises when the electron-electron dipolar reservoir establishes thermal contact with the nuclear Zeeman reservoir. This takes place when the characteristic electronic resonance line width is of the order of the nuclear Larmor frequency. Electron-electron cross relaxation between spins with difference in energy equal

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to the nuclear Zeeman energy is absorbed or emitted by the electronic dipolar reservoir, changing its spin temperature and the nuclear polarisation is enhanced. For thermal mixing both the forbidden and the allowed transitions can be involved.

In the first embodiment where the polarising agent is an OMRI contrast agent, the method may be conveniently carried out by using a first magnet for providing the polarising magnetic field and a second magnet for providing the primary magnetic field for MR imaging. The same magnet could be used for both purposes. Figure 1 of the accompanying drawings is a schematic representation of an apparatus suitable for carrying out the first embodiment of the invention. A freestanding polarising magnet (1) optionally together with a filter surrounds an EPR resonator (2) which provides the nuclear polarisation. A container (3) comprising a pump is provided for carrying the contrast composition which is delivered to a subject (4) by a delivery line (5). The subject is situated within a conventional MR scanner (6).

In the above apparatus, a dielectric resonator may be used in the dynamic nuclear polarisation process. Generally speaking, dynamic nuclear polarisation requires a volume with a fairly strong high frequency magnetic field and an accompanying electric field which is made as small as possible. A dielectric resonator may be used to provide a preferred field arrangement in which the magnetic field lines are shaped like a straw in a sheaf of corn with an electric field forming circles like the thread binding the sheaf. A field arrangement of this type may be formed by one of several rings or tubes of a material with a high dielectric constant and low loss. The man skilled in the art will appreciate that such a tube will exhibit different electromagnetic resonant modes. One of the dominant modes has the desired characteristic of electric field

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circulating around the tube axis within the wall and being zero at the axis and everywhere perpendicular to it. The magnetic field on the other hand is concentrated around the tube axis and mainly directed along it. The composition to be polarised is conveniently placed inside the resonator which is itself placed inside a metal box with a clearance typically of the order of the size of the resonator, and is excited to the desired resonance with a coupling loop or the like. The metal box ensures that the electromagnetic energy does not leak away by radiation. Figure 2 of the accompanying drawings shows a dielectric resonator (1) (with an axis of rotational symmetry (2)) within a metal box (3).

An alternative to the dielectric resonator is a resonant cavity of which several are known to those skilled in the art. One simple and efficient resonant cavity is a metal box, such as a cylindrical metal box. A suitable mode is the one known as TM_{1,1,0} which produces a perpendicular magnetic field on the axis of the cavity. It is possible to excite two such modes in the same cavity at the same frequency producing fields which are mutually perpendicular. By arranging them to have a 90° phase difference a rotating field can be produced which is especially efficient for implementing dynamic polarisation with a minimum of dissipation in the sample. Modes with similar field distributions for different shapes of cavities e.g. rectangular cavities are familiar to those skilled in the art.

The composition may also be dispersed into a plurality of compartments during the dynamic nuclear polarisation step. Thus the composition might be typically divided into parallel channels provided, for example, by parallel separating plates, discs or tubes, typically open-ended tubes. The electric losses (eddy currents) in the composition caused by the magnetic field are decreased by dividing the composition into

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smaller volumes using electrically isolating barriers, preferably situated perpendicular to the field. If the composition is in a cylindrical vessel surrounded by a dielectric resonator as described hereinbefore, the isolating barriers would be planes passing radially from the vessel axis to its wall. A simpler and more practical arrangement is to polarise the composition in a container which contains a plurality of thin-walled tubes of an isolating material such as quartz, glass or plastic. This has the advantage of reducing the electric losses in the composition which allows a larger volume of composition to be polarised for the same applied electromagnetic power. The walls, the inner, outer or both of the tubes may similarly serve as the substrate onto which the OMRI contrast agent is bound so that pressure applied to one end of the container may force the polarized, substantially OMRI contrast agent free, fluid high T_1 agent from the container, for example with a delivery line leading to the subject (patient) undergoing MR examination.

It is envisaged that in the first embodiment of the method according to the invention, use may be made of any known OMRI contrast agent capable of polarising a high T_1 agent to an extent such that a diagnostically effective contrast enhancement, in the sample to which the high T_1 agent is administered, is achieved. Where the OMRI contrast agent is a paramagnetic free radical, the radical may be conveniently prepared *in situ* from a stable radical precursor by a conventional physical or chemical radical generation step shortly before polarisation, or alternatively by the use of ionising radiation. This is particularly important where the radical has a short half-life. In these cases, the radical will normally be non-reusable and may conveniently be discarded once the separation step of the method according to the invention has been completed.

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In solids, it is preferred to effect dynamic nuclear polarisation by irradiating an electron spin at low temperature and high field. Specific examples of dynamic nuclear polarisation of solid high T_1 agents are:

(1) ^{15}N -Ala labelled T4-lysosome and ^{13}C -Glycine in frozen aqueous solutions of 60:40 glycerol/water with the free radical 4-amino TEMPO as the source of electron polarisation (D.A. Hall, D. Maus, G. Gerfen and R.G. Griffin, Science, 1997), Enhancements of ca. 50 and 100 were obtained, respectively, at 5T and 40K;

(2) Carboxy- ^{13}C labelled glycine in frozen aqueous solution of 60:40 glycerol/water with TEMPO as the free radical. An enhancement of 185 at 5T and 14K was obtained (G.J. Gerfen, L.R. Becerra, D.A. Hall, R.G. Griffin, R.J. Temkin, D.J. Singel, J. Chem. Phys. 102(24), 9494-9497 (1995);

(3) Dynamic polarisation of protons and deuterons in 1,2-ethanediol doped with complexes of Cr at 2.5T. The obtained degree of polarisation is 80% (W. De Boer and T.O Niinikoski, Nucl. Instrum. Meth. 114, 495 (1974)).

Preferably of course a chosen OMRI contrast agent will exhibit a long half-life (preferably at least one hour), long relaxation times (T_{1e} and T_{2e}), high relaxivity and a small number of ESR transition lines. Thus the paramagnetic oxygen-based, sulphur-based or carbon-based organic free radicals or magnetic particles referred to in WO-A-88/10419, WO-A-90/00904, WO-A-91/12024, WO-A-93/02711 or WO-A-96/39367 would be suitable OMRI contrast agents.

However, OMRI contrast agents useful in the first embodiment of the present method are not limited to paramagnetic organic free radicals. Particles exhibiting the magnetic properties of paramagnetism, superparamagnetism, ferromagnetism or ferrimagnetism may also be useful OMRI contrast agents, as may be other particles having associated free electrons.

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Superparamagnetic nanoparticles (eg. iron or iron oxide nanoparticles) may be particularly useful. Magnetic particles have the advantages over organic free radicals of high stability and a strong electronic/nuclear spin coupling (i.e. high relaxivity) leading to greater Overhauser enhancement factors.

For the purposes of administration, the high T₁ agent should be preferably administered in the absence of the whole of, or substantially the whole of, the OMRI contrast agent. Preferably at least 80% of the OMRI contrast agent is removed, particularly preferably 90% or more, especially preferably 95% or more, most especially 99% or more. In general, it is desirable to remove as much OMRI contrast agent as possible prior to administration to improve physiological tolerability and to increase T₁. Thus preferred OMRI contrast agents for use in the first embodiment of the method according to the invention are those which can be conveniently and rapidly separated from the polarised high T₁ MR imaging agent using known techniques as discussed below. However where the OMRI contrast agent is non-toxic, the separation step may be omitted. A solid (eg. frozen) composition comprising an OMRI contrast agent and a high T₁ agent which has been subjected to polarisation may be rapidly dissolved in saline (eg. warm saline) and the mixture injected shortly thereafter.

In the separation step of the first embodiment of the method of the invention, it is desirable to remove substantially the whole of the OMRI contrast agent from the composition (or at least to reduce it to physiologically tolerable levels) as rapidly as possible. Many physical and chemical separation or extraction techniques are known in the art and may be employed to effect rapid and efficient separation of the OMRI contrast agent and high T₁ agent. Clearly the more preferred separation techniques are those which can be effected rapidly and particularly those which allow

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separation in less than one second. In this respect, magnetic particles (eg. superparamagnetic particles) may be advantageously used as the OMRI contrast agent as it will be possible to make use of the inherent magnetic properties of the particles to achieve rapid separation by known techniques. Similarly, where the OMRI contrast agent or the particle is bound to a solid bead, it may be conveniently separated from the liquid (i.e. if the solid bead is magnetic by an appropriately applied magnetic field).

For ease of separation of the OMRI contrast agent and the high T_1 agent, it is particularly preferred that the combination of the two be a heterogeneous system, eg. a two phase liquid, a solid in liquid suspension or a relatively high surface area solid substrate within a liquid, eg. a solid in the form of beads fibres or sheets disposed within a liquid phase high T_1 agent. In all cases, the diffusion distance between the high T_1 agent and OMRI contrast agent must be small enough to achieve an effective Overhauser enhancement. Certain OMRI contrast agents are inherently particulate in nature, eg. the paramagnetic particles and superparamagnetic agents referred to above. Others may be immobilized on, absorbed in or coupled to a solid substrate or support (eg. an organic polymer or inorganic matrix such as a zeolite or a silicon material) by conventional means. Strong covalent binding between OMRI contrast agent and solid substrate or support will, in general, limit the effectiveness of the agent in achieving the desired Overhauser effect and so it is preferred that the binding, if any, between the OMRI contrast agent and the solid support or substrate is weak so that the OMRI contrast agent is still capable of free rotation. The OMRI contrast agent may be bound to a water insoluble substrate/support prior to the polarisation or the OMRI contrast agent may be attached/bound to the substrate/support after

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polarisation. The OMRI contrast agent may then be separated from the high T_1 agent e.g. by filtration before administration. The OMRI contrast agent may also be bound to a water soluble macromolecule and the OMRI contrast agent-macromolecule may be separated from the high T_1 agent before administration.

Where the combination of an OMRI contrast agent and high T_1 agent is a heterogeneous system, it will be possible to use the different physical properties of the phases to carry out separation by conventional techniques. For example, where one phase is aqueous and the other non-aqueous (solid or liquid) it may be possible to simply decant one phase from the other. Alternatively, where the OMRI contrast agent is a solid or solid substrate (eg. a bead) suspended in a liquid high T_1 agent the solid may be separated from the liquid by conventional means eg. filtration, gravimetric, chromatographic or centrifugal means. It is also envisaged that the OMRI contrast agents may comprise lipophilic moieties and so be separated from the high T_1 agent by passage over or through a fixed lipophilic medium or the OMRI contrast agent may be chemically bound to a lipophilic solid bead. The high T_1 agent may also be in a solid (eg. frozen) state during polarisation and in close contact with a solid OMRI contrast agent. After polarisation it may be dissolved in heated water or saline or melted and removed or separated from the OMRI contrast agent where the latter may be toxic and cannot be administered.

One separation technique makes use of a cation exchange polymer and a cationic OMRI contrast agent, eg. a triarylmethyl radical carrying pendant carboxylate groups. Alternatively acidifying the solution to around pH 4 may cause the OMRI contrast agent to precipitate out. Separation may then be carried out for example by filtration followed by neutralisation. An alternative technique involves adding ions which causes

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precipitation of ionic OMRI agents which may then be filtered off.

Certain OMRI contrast agents, such as the triarylmethyl radical, may have an affinity for proteins. Thus, after polarisation, a composition containing an OMRI contrast agent with a protein affinity may be passed through or over a protein in a form which exposes a large surface area to the agent eg. in particulate or surface bound form. In this way, binding of the OMRI contrast agent to the protein enables it to be removed from the composition.

Alternatively when a hydrophilic high T_1 agent is in a solid (eg. frozen) form it may be brought into contact with a hydrophobic OMRI contrast agent which is dissolved in an organic fluid with a melting temperature higher than the high T_1 agent. The mixture is frozen and polarisation performed. After polarisation, the mixture is heated and the solid OMRI contrast agent and its solvent are removed. The high T_1 agent will remain hyperpolarised for a significant time in the frozen state and may be transported long distances before being dissolved in water or saline for injection.

In a second embodiment of the method according to the invention, hyperpolarisation of the nuclei is effected by a hyperpolarisable gas. In this second embodiment, step (i) of the method according to the invention comprises:

(a) hyperpolarising a hyperpolarisable gas before, during or after introducing a high T_1 agent thereto whereby to cause nuclear polarization of said high T_1 agent;

(b) dissolving in a physiologically tolerable solvent said high T_1 agent.

By hyperpolarisable gas is meant a gas with a non-zero spin angular momentum capable of undergoing an electron transition to an excited electron state and thereafter of decaying back to the ground state.

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Depending on the transition that is optically pumped and the helicity of the light a positive or negative spin hyperpolarisation may be achieved (up to 100%).

Examples of gases suitable for use in the second embodiment of the method of the invention include the noble gases He (eg. ³He or ⁴He) and Xe (eg. ¹²⁹Xe), preferably He, particularly preferably ³He. Alkali metal vapours may also be used eg. Na, K, Rb, Cs vapours.

Mixtures of the gases may also be used or the hyperpolarisable gas may be used in liquid or solid form. The term hyperpolarisable gas also covers any gas with non-zero nuclear spin which may be polarised by optical pumping and is preferably ¹²⁹Xe or ³He.

It will be appreciated that in the second embodiment of the invention, the hyperpolarised gas may transfer polarisation to the nuclear spin system of a high T₁ agent directly or indirectly. Where the high T₁ agent is to be polarised indirectly by water vapour, it may be advantageously water soluble.

For the purposes of polarisation according to the second embodiment of the invention, the high T₁ agent may be generally in gaseous, liquid or solid form.

Where the high T₁ agent is polarised whilst in a gaseous state, it is convenient (for the purposes of separation from the hyperpolarised gas and of administration) to be able to rapidly convert it into a liquid or solid. This has the added benefit of significantly increasing T₁. Thus removing the elevated pressure and temperature imposed on the gas mixture will lead to rapid cooling and condensation. Yet further cooling is possible by, for example, contacting the polarised high T₁ agent with a cold surface.

In a preferred embodiment, a hyperpolarised fluid eg. ¹²⁹Xe at elevated pressure and/or low temperature is passed through a column of solid ¹³C enriched and/or ¹⁹F enriched high T₁ agent until steady state polarisation of the solid is almost achieved. In general any of the

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above-mentioned ^{13}C enriched agents may be used.

In another preferred embodiment, a hyperpolarised gas is frozen/crystallised on the solid/frozen surface of a solid high T_1 agent which has been prepared with as large a surface area as possible. The mixture may be transported before warm administrable media (eg. saline) is added and physiological temperature reached before injection.

^{129}Xe gas can be produced in a highly spin polarised state in macroscopic quantities. Due to the limited solubility and inert nature of xenon there is interest in transferring the polarisation to other nuclei.

It can also be produced by irradiating a polarising agent, e.g. with an electron spin resonance transition stimulating radiation (e.g. microwave radiation). This forms a further aspect of the invention. Viewed from this aspect the invention provides a method of magnetic resonance investigation of a sample, preferably of a human or non-human animal body, said method comprising:

i) producing solid hyperpolarised ^{129}Xe by irradiating a polarising agent whereby to cause dynamic nuclear polarisation.

In the above method said polarising agent is preferably a substance containing an unpaired electron, for example nitroxide, trityl, Cr(V), or the OMRI agents mentioned above.

Considerable interest has been generated in the novel technique of MR lung imaging using hyperpolarised gases such as ^3He and ^{129}Xe as inhaled contrast media. However, the production of these gases in their hyperpolarised form is labourious and time consuming. At the present time, ^3He , where most of the interest is today, can be generated at a rate of a few liters an hour. However, if the hyperpolarisation could be done in the liquid or solid phase, much higher production rates would be possible. Using only "brute force", i.e. milliKelvin temperatures and >10 T, fields would be an

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extremely costly method, however, "double brute force", i.e. irradiation of frozen Xe in the presence of a free radical (metal ion, trityl radical, nitroxide, etc.) at a comparatively moderate temperature (a few K) would be a more practical method. The radical would be added either in pure form or bound to a matrix. After the irradiation had been carried out, heating of the sample would release the hyperpolarised gas and a new batch of Xe could be condensed and irradiated. Since the hyperpolarisation in this case is carried out on solid Xe, the possibilities of producing large amounts of gas would be considerable.

The main relaxation mechanism for solid ^{129}Xe is spin exchange with the rapidly relaxing ^{131}Xe , the major component in natural xenon. The magnetogyric ratio of ^{129}Xe and ^{131}Xe differs by a factor of four. Normally the line widths of the resonances of solids are on the order of a few kHz. When the difference in Larmor frequency is on the same order as the line width, the polarisation of the nuclei will rapidly equilibrate. Assuming that we have a cold (colder than the freezing point of xenon, around 150 K, depending on the pressure), finely divided (some micrometers grain size), sample of a ^{13}C labelled substance with a long T_1 in the solid and allow hyperpolarised xenon to form frost on the powder. If this operation is performed in a magnetic field of suitable strength the ^{129}Xe and the ^{13}C will overlap and Xe-C spin flip-flops will be efficient, equilibrating the polarisation between xenon and carbon. The xenon can then be pumped off and the process repeated until a suitable level of polarisation is achieved. What the suitable field strength depends on the exact lineshapes but assuming line widths on the order of 5-10 kHz, which is quite normal for solids, the optimum field is around 10 mT, typically the field on the outside of an NMR-magnet or a small toy magnet. The basis for this is that the centre frequency of the line is field

dependent whereas the linewidth is essentially independent of the field.

Figure 3 shows the behaviour of such a system at various field strengths. One important factor to take into account is that all the nuclei in the sample must be taken into consideration. This method will work for transfer from ^{129}Xe to ^{13}C and possibly to ^{29}Si but it is not expected to work with ^{15}N which has a resonance frequency that is closer to ^{131}Xe than to ^{129}Xe . There will be interference from quadrupolar nuclei like ^{23}Na , ^{79}Br , ^{81}Br , ^{127}I and a number of transition metals, all having resonance frequencies similar to carbon.

In order to generate a hyperpolarised gas, the gas is first subjected to a discharge or other means of excitation (eg. an appropriate radiofrequency) which creates a metastable unpaired electron spin state and is then optically (eg. laser) pumped at an appropriate frequency to create electron hyperpolarisation. The various methods for achieving this are well known to those skilled in the art or are described in *inter alia* US-A-5545396.

Preferred hyperpolarisable gases for use in the second embodiment of the method according to the invention are those which can be conveniently and rapidly separated from the polarised high T_1 agent. Noble gases are particularly useful given their very low boiling points and inertness. Preferably the chosen gas will exhibit a long hyperpolarisability half-life (preferably at least 1000s, particularly preferably at least 4000s and especially preferably 8000s or more).

A hyperpolarised gas may, if desired, be stored for extended periods of time in a hyperpolarised state. This is achieved by maintaining the gas at very low temperatures, preferably in a frozen state.

For ease of separation of the hyperpolarisable gas and the high T_1 agent, the combination of the two may be advantageously a heterogeneous system, eg. the high T_1

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agent is a solid at ambient temperatures. In all cases, the diffusion distance between the high T_1 agent and gas, fluid or solid must be small enough to achieve an effective polarisation.

In the separation step of the second embodiment of the method of the invention, it is desirable to remove substantially the whole of the hyperpolarisable gas from the composition (or at least to reduce it to physiologically tolerable levels) as rapidly as possible. If desired, the gas may be reused which may be an important consideration given the expense of noble gases. Many physical and chemical separation or extraction techniques known in the art may be employed to effect rapid and efficient separation of the hyperpolarisable gas and high T_1 agent. Clearly the more preferred separation techniques are those which can be effected rapidly and particularly those which allow separation in a fraction of the relaxation time T_1 of the high T_1 agent.

In a third embodiment of the method of the invention, hyperpolarisation of the MR imaging nuclei is effected by the use of a high field as described in US-A-5479925 (GEC) and US-A-5617859 (GEC). US-A-5479925 discloses a method for generating MR angiograms in which a contrast agent is passed through a small, high field polarising magnet *ex vivo*, in order to generate a high longitudinal magnetisation in the agent prior to its administration to the subject. There is however no mention or suggestion of the use of high T_1 agents to achieve an improved effect.

Generally speaking, polarisation of an MR imaging nuclei may be achieved by thermodynamic equilibration at low temperature and high magnetic field. Where the contrast medium to be administered is a solid material (e.g. crystalline), it may be introduced into a magnetic field at very low temperature. Under these conditions, T_1 is very long (typically many hours or months) and

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consequently it takes an unacceptably long time for the medium to reach thermodynamic equilibrium. Thus if the contrast medium undergoes small movements in the gradient field for example by exposure to a magnetic field gradient and ultrasound or by relative movement within the gradient field, T_1 will drop. When thermodynamic equilibrium is attained, all nuclei in the contrast medium will be highly polarised relative to room temperature and to normal magnetic fields used in MRI. This procedure has the advantage of allowing the contrast medium to be removed from the magnet and transported in a "ready-to-use" form to the place where it is to be used. Preferably but not essentially transport may take place at a relatively low temperature (e.g. at liquid nitrogen temperature). The T_1 of the high T_1 solid contrast medium will be long enough to allow transport at ambient temperature before use.

One of the main obstacles in using so-called 'brute force' polarisation as a method for hyperpolarising samples are the long T_1 values at low temperatures and high fields, typically several weeks at temperatures below 1 K. However it has been found that it is possible to utilise the non-linear field dependence of T_1 to shorten the time necessary for relaxation by a gradual increase of the external magnetic field.

As stated above, it is of great interest to obtain hyperpolarised injectable contrast agents. Theoretically, the simplest way of obtaining a highly spin-polarised material is to cool it to a very low temperature in a strong magnetic field and let the sample reach thermal equilibrium. The major practical problem in using this technique is the time required for the thermal equilibration to occur. At temperatures below 1 K the time constant for that process, T_1 , might be on the order of weeks.

The time constant of nuclear longitudinal relaxation, T_1 , shows a quadratic dependence on the field

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strength in solid materials:

$$T_1 = T_{1,0} + cB^2$$

Where $T_{1,0}$ is the time constant for relaxation at no external magnetic field, c is a constant, and B is the external magnetic field.

The rate of magnetisation of the sample, dM/dt , at a given field strength will then be given by:

$$dM/dt = (M_{\max} - M) / T_1$$

Where M_{\max} is the magnetisation of the sample after complete relaxation at the final field. Since the field-dependence of the time constant is non-linear, it will be possible to obtain a larger magnetisation at a given time by constantly tuning the external magnetic field so that the rate of magnetisation all the time is as big as possible. The example shown in attached Figures 4 and 5 was chosen to simulate the behaviour of the carbonyl carbon in solid sodium acetate. The T_1 at 7 T is 1700 seconds and the $T_{1,0}$ is about 5 seconds. The time to reach the same degree of magnetisation as after 1700 seconds at a constant field of 7 T, is reduced to 1390 seconds, a reduction of almost 20%, which could easily reduce the equilibration time by one week at milliKelvin temperatures. The optimised field-ramp is shown in Figure 4, whilst Figure 5 shows the expected values from a numerical integration of the equation for dM/dt given above. This process will be applicable for all nuclei with spin but will be most interesting with compounds with long T_1 values

As stated above, one of the main obstacles in using so-called 'brute force' polarisation as a method for hyperpolarising samples is the long T_1 values found at low temperatures and high fields, typically several weeks at temperatures below 1 K. It is possible to use

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the technique of low-field matching to increase the relaxation rate and the degree of polarisation of the nuclear spins in solids at low temperature. This has the additional advantage that a brute force polariser does not need to possess any radio frequency electronics.

It is well known that different nuclei in the same molecule will relax with different time constants. A way of speeding up the polarisation of the interesting ^{13}C nucleus and at the same time obtaining a better polarisation is to use cross-polarisation from the quickly relaxing proton to the slowly relaxing carbon, a method routinely used in solid-state NMR spectroscopy. Due to the big difference in magnetogyric ratio between the proton and ^{13}C , the energy difference is large and hence the polarisation transfer slow. The magnetogyric ratio of the proton is roughly a factor of four larger than that of carbon. The situation can be improved by utilising the procedure of spin locking under Hartman-Hahn conditions. Spin-lock (90_x -long pulse, y) at both nuclei with the amplitude (B_1) of the long pulse satisfying the Hartman-Hahn condition:

$$\gamma H B_{1H} = \gamma C B_{1C}$$

where γH is the magnetogyric ratio of hydrogen, γC is the magnetogyric ratio of carbon, B_{1H} is the proton excitation field and B_{1C} is the carbon excitation field.

This allows for mutually matched flip-flops of the spins. Since this is a spin-spin process, it usually occurs on time scales from about $100\ \mu\text{s}$ to a few ms.

One problem with this is that radiofrequency electronics are required and furthermore the homogeneity of the magnetic field must be high enough to allow precise pulse angles. A way to circumvent this problem is the following.

A crude way of stating the Hartman-Hahn condition

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is to say that spin diffusion is efficient when the resonance lines of the two nuclei overlap. Assume the substrate to be a solid material with a half-height line width of 5 kHz. This linewidth is caused by dipolar coupling and is independent of the external field. The Hartman-Hahn condition is now restated as follows. Efficient spin-diffusion takes place when the maxima of the two resonances are separated by less than the sum of their half-height line-widths. The field where this condition is fulfilled is derived as follows.

The resonance frequency, ν is given by:

$$\nu = \gamma B_0 / 2\pi \quad (1)$$

where γ is the magnetogyric ratio, and B_0 is the external magnetic field. The required separation, ν , was 5 kHz:

$$\nu = \nu_H - \nu_C = 5000 \text{ s}^{-1} \quad (2)$$

Combination of equation (1) and (2) gives:

$$\nu = B_0 (\gamma_H - \gamma_C) / 2\pi$$

which can be rewritten as:

$$B_0 = \nu 2\pi / (\gamma_H - \gamma_C) = 156 \mu\text{T}$$

This field is within a factor 3 of the earth's magnetic field which means that if the sample is removed from the polarising magnet for a few seconds the polarisation will equilibrate between carbon and hydrogen on a time scale similar to T_2 so there will be plenty of time to put the sample back into the magnet again before the T_1 relaxation becomes significant, even if it has to be kept in mind that the T_1 shortens dramatically for solids in low fields. It will, however, never be as short as the T_2 .

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This procedure can be repeated after the protons have repolarised again, successively building up the carbon polarisation until the spin-temperature of the two nuclei become the same. The protons in solid sodium acetate has, at room temperature, a T_1 of 31 s whilst the T_1 of the carbonyl carbon in the same sample is 1700 s. If this difference could be used completely there would be a shortening of the polarisation time by a factor of 55. Generally fluorine relaxes even faster than protons and it would be possible to include a fluorine atom in the contrast agent molecule as an internal relaxation agent.

It is also possible to use quadrupolar nuclei for this process. The sodium ions in solid sodium acetate have a T_1 of 1.7 s at room temperature. Sodium has a magnetogyric ratio only slightly higher than carbon which means that equilibration occurs at a much higher field, in this case at 8.9 mT, a field found about 15 cm above the Dewar of a 7 T NMR magnet. This is of great help for rapid polarisation of samples at low temperature, but is also a problem in the storage of polarised samples. The magnetic storage field must be large enough to avoid overlap of the resonances of the wanted nucleus and any rapidly relaxing quadrupolar nuclei. It is known that this phenomenon causes the rapid relaxation of frozen ^{129}Xe in a low field, when spin diffusion to the quadrupolar ^{131}Xe becomes efficient.

There is also the possibility of transferring polarisation from unpaired electrons to carbon. Due to the great difference in magnetogyric ratios this requires a field much lower than the earth magnetic field to become efficient. Such a low field requires that the sample be moved to a magnetically shielded area. One way of achieving this is to have a small magnet with opposite polarity some distance along the polarity axis of the main coil. With careful design the

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fields can be made to cancel in the centre of the small magnet.

The magnetic field strength used in this third embodiment of the invention should be as high as possible, preferably >1T, more preferably 5T or more, especially preferably 15T or more. The temperature should be very low e.g. 100K or less, preferably 4.2K or less, more preferably 1K or less, even more preferably 0.1K or less, especially preferably 1mK or less.

Thus viewed from a further aspect the present invention provides a method for preparing polarised high T_1 agents, said method comprising:

(i) subjecting a high T_1 agent to a high magnetic field (e.g. 1T or more) at low temperature (e.g. 100K or less);

(ii) exposing the agent to a T_1 shortening effect in order to attain thermodynamic equilibrium at said low temperature.

The T_1 shortening effect may be provided by exposure to a variable magnetic field gradient but it may also be achieved by adding magnetic material (e.g. paramagnetic, superparamagnetic or ferromagnetic materials) to the agent during the period when the agent is exposed to low temperature, field cycling to a field allowing cross polarisation, gradually increasing the magnetic field at such a rate that the increase in polarisation of the high T_1 agent is maximised, gradually decreasing the temperature at such a rate that the increase in polarisation of the high T_1 agent is maximised, or adding a material with unpaired electrons during the period when the high T_1 agent is exposed to said low temperature. Possible T_1 shortening agents include Gd and NO[•] but preferred T_1 shortening agents are O₂ and NO which may be conveniently separated from the high T_1 agent before transportation and subsequent use.

In the third embodiment of the invention, both the high T_1 agent and the aqueous solvent (eg. water) in

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which it is dissolved may be polarised. This may be carried out at low temperature conveniently in the same magnetic field and after mixing the adminstrable composition should be warmed very rapidly prior to administration.

Thus viewed from a further aspect, the present invention provides an administrable composition comprising a polarised high T₁ agent and polarised water.

The high T₁ agents used in the method according to the invention may be conveniently formulated with conventional pharmaceutical or veterinary carriers or excipients. Formulations manufactured or used according to this invention may contain, besides the high T₁ agent, formulation aids such as are conventional for therapeutic and diagnostic compositions in human or veterinary medicine. Thus the formulation may for example include stabilizers, antioxidants, osmolality adjusting agents, solubilizing agents, emulsifiers, viscosity enhancers, buffers, etc. The formulation may be in forms suitable for parenteral (eg. intravenous or intraarterial) or enteral (eg. oral or rectal) application, for example for application directly into body cavities having external voidance ducts (such as the lungs, the gastrointestinal tract, the bladder and the uterus), or for injection or infusion into the cardiovascular system. However solutions, suspensions and dispersions in physiological tolerable carriers will generally be preferred.

For use in *in vivo* imaging, the formulation, which preferably will be substantially isotonic, may conveniently be administered at a concentration sufficient to yield a 1 micromolar to 10M concentration of the high T₁ agent in the imaging zone; however the precise concentration and dosage will of course depend upon a range of factors such as toxicity, the organ targeting ability of the high T₁ agent and the administration route. The optimum concentration for the

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MR imaging agent represents a balance between various factors. In general, optimum concentrations would in most cases lie in the range 0.1mM to 10M, preferably more than 10mM, especially more than 100mM. Isotonic solution may be especially preferred. In certain circumstances concentrations above 1M are preferred. Formulations for intravenous or intraarterial administration would preferably contain the high T₁ agent in concentrations of 10mM to 10M, especially more than 50mM. For bolus injection the concentration may conveniently be 0.1mM to 56M, preferably more than 200mM, more preferably more than 500mM. In certain circumstances, the preferred concentration is above 1M, even more preferably above 5M.

PARENTERALLY ADMINISTRABLE FORMS SHOULD OF COURSE BE STERILE AND FREE FROM PHYSIOLOGICALLY UNACCEPTABLE AGENTS, AND SHOULD HAVE LOW OSMOLALITY TO MINIMIZE IRRITATION OR OTHER ADVERSE EFFECTS UPON ADMINISTRATION AND THUS THE FORMULATION SHOULD PREFERABLY BE ISOTONIC OR SLIGHTLY HYPERTONIC. SUITABLE VEHICLES INCLUDE AQUEOUS VEHICLES CUSTOMARILY USED FOR ADMINISTERING PARENTERAL SOLUTIONS SUCH AS SODIUM CHLORIDE SOLUTION, RINGER'S SOLUTION, DEXTROSE SOLUTION, DEXTROSE AND SODIUM CHLORIDE SOLUTION, LACTATED RINGER'S SOLUTION AND OTHER SOLUTIONS SUCH AS ARE DESCRIBED IN REMINGTON'S PHARMACEUTICAL SCIENCES, 15TH ED., EASTON: MACK PUBLISHING CO., PP. 1405-1412 AND 1461-1487 (1975) AND THE NATIONAL FORMULARY XIV, 14TH ED. WASHINGTON: AMERICAN PHARMACEUTICAL ASSOCIATION (1975). THE COMPOSITIONS CAN CONTAIN PRESERVATIVES, ANTIMICROBIAL AGENTS, BUFFERS AND ANTIOXIDANTS CONVENTIONALLY USED FOR PARENTERAL SOLUTIONS, EXCIPIENTS AND OTHER ADDITIVES WHICH ARE COMPATIBLE WITH THE HIGH T₁ AGENTS AND WHICH WILL NOT INTERFERE WITH THE MANUFACTURE, STORAGE OR USE OF THE PRODUCTS.

WHERE THE HIGH T₁ AGENT IS TO BE INJECTED, IT MAY BE CONVENIENT TO INJECT SIMULTANEOUSLY AT A SERIES OF

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administration sites such that a greater proportion of the vascular tree may be visualized before the polarization is lost through relaxation.

The dosages of the high T₁ agent used according to the method of the present invention will vary according to the precise nature of the high T₁ agents used, of the tissue or organ of interest and of the measuring apparatus. Preferably the dosage should be kept as low as possible while still achieving a detectable contrast effect. In general, the maximum dosage will depend on toxicity constraints.

The contents of all publications referred to herein are hereby incorporated by reference.

The invention is illustrated with reference to the following non-limiting Examples and the accompanying drawings in which:

Figure 1 is a schematic representation of an apparatus suitable for carrying out the first embodiment of the invention;

Figure 2 shows a dielectric resonator (1) (with an axis of rotational symmetry (2)) within a metal box (3);

Figure 3 shows the behaviour of a system at various field strengths;

Figure 4 shows a build-up of magnetisation versus time; and

Figure 5 shows magnetic field versus time.

Example 1

A high T₁ agent is placed in a chamber at very low temperature (about 4K). Fluient O₂ is added and crystallised on the surface of the high T₁ agent. In a separate chamber, frozen H₂O is subjected to the same treatment as the high T₁ agent. Both chambers are placed in a strong magnetic field (about 15T) and the

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temperature kept low.

When thermodynamic equilibrium is reached, the temperature is increased to about 200K. The oxygen disappears as a gas. The high T₁ agent and the frozen H₂O are mixed and stored until needed. The temperature is increased and the solution comprising polarised high T₁ agent and hyperpolarised water is injected.

Example 2

300 mg of sterile Na₂¹³CO₃ or NaH¹³CO₃ is placed inside a 10 ml plastic injection syringe. The gas inside the syringe is enriched with >20% oxygen. The syringe is placed inside a magnet (1-20T) at a temperature of about 4K (0.001-5K) until thermodynamic equilibrium is reached.

The syringe is removed and transported to the subject located in the MRI magnet. 10 ml of sterile Ringers Solution (at 37°C, pH 7.4) is aspirated and injected at a rate of 10 ml/sec immediately after the high T₁ agent has dissolved. ¹³C MRI is performed using a fast pulse sequence. T₁ in the blood is about 20s and the distribution of the agent is followed on the MR imager.

Example 3

To a sample of sodium acetate (1-¹³C) is added α,γ-bisphenyl-β-phenylallyl benzene complex (5% w/w). The compounds are milled together to give an intimate mixture, which is transferred to a borosilicate glass ampule. This is then repeatedly evacuated and filled with helium. The last time a pressure of a 200 mbar of helium is left in the ampule, which is then flame sealed.

The sample is polarized by microwaves (70 GHz) for

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at least one hour at a field of 2.5 T at a temperature of 4.2 K. The progress of the polarization process is followed by in situ NMR (fast adiabatic passage). When a suitable level of polarization has been reached, the ampule is rapidly removed from the polarizer and, while handled in a magnetic field of no less than 50 mT, cracked open and the contents are quickly discharged and dissolved in warm (40°C) water.

Experiment 1: This solution is quickly transferred to a spectrometer and ¹³C spectrum with enhanced intensity is recorded.

Experiment 2: The sample solution is inserted into an MRI machine with ¹³C capability and a picture with enhanced intensity and contrast is obtained by a single shot technique.

Experiment 3: The solution is quickly injected into a rat and a ¹³C MRI picture with enhanced intensity and contrast is obtained, also in this case, by utilization of a single shot technique.

Example 4

To a sample of sodium bicarbonate -¹³C is added MnCl₂ (5% w/w). The compounds are milled together to give an intimate mixture, which is transferred to a borosilicate glass ampule. This is then repeatedly evacuated and filled with helium. The last time a pressure of a 200 mbar of helium is left in the ampule, which is then flame sealed.

The sample is polarized by microwaves (70 GHz) for at least 1 hour at a field of 2.5 T at a temperature of 4.2 K. The progress of the polarization process is followed by in situ NMR (fast adiabatic passage). When a suitable level of polarization has been reached, the ampule is rapidly removed from the polarizer and, while handled in a magnetic field of no less than 50 mT, cracked open and the contents are quickly discharged and

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dissolved in warm (40°C) water.

Experiment 1: This solution is quickly transferred to a spectrometer and ¹³C spectrum with enhanced intensity is recorded.

Experiment 2: The sample solution is inserted into an MRI machine with ¹³C capability and a picture with enhanced intensity and contrast is obtained by a single shot technique.

Experiment 3: The solution is quickly injected into a rat and a ¹³C MRI picture with enhanced intensity and contrast is obtained, also in this case, by utilization of a single shot technique.

Examples 5-7

Low-field pumping of ¹³C

Example 5

A sample of solid 1-¹³C-2,2,2',2',2",2"-hexadeuterotris(hydroxymethyl)nitromethane was subjected to a magnetic field of 6.56 T at a temperature of 2.5 K for 10 minutes. The sample was then removed from the centre of the magnet to the stray field (7 mT) for a duration 1 s and then returned to the magnet. After another 10 minutes the process was repeated one more. An ¹³C-NMR spectrum of the solid sample was recorded and the signal was found to be in accordance with thermal equilibrium at 6.56 T and 2.5 K.

T₁-values for the ¹³C atom in 1-¹³C-2,2,2',2',2",2"-hexadeuterotris(hydroxymethyl)nitromethane

H ₂ O, air saturated, 37°C, 7 T	95 s
H ₂ O, degassed, 37°C, 7 T	102 s
Human bloodplasma, 37°C, 7 T	60 s
Solid, 20°C, 7 T	22 min

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Solid, -30°C, 7 T	47 min
Solid, 2.5 K, 7 T	55 hours

Example 6

A sample of solid 1-¹³C-1,1-bis(hydroxydideuteromethyl)-2,2,3,3-tetradeuterocyclopropane was subjected to a magnetic field of 6.56 T at a temperature of 2.5 K for 10 minutes. The sample was then removed from the centre of the magnet to the stray field (7 mT) for a duration of 1 s and then returned to the magnet. After another 10 minutes the process was repeated one more. A ¹³C-NMR spectrum of the solid sample was recorded and the signal was found to be in accordance with thermal equilibrium at 6.56 T and 2.5 K.

Example 7

A sample of solid 2-¹³C-2,2-bis(trideuteromethyl)-1,1,3,3-tetradeuteropropane-1,3-diol was subjected to a magnetic field of 6.56 T at a temperature of 2.5 K for 10 minutes. The sample was then removed from the centre of the magnet to the stray field (7 mT) for a duration 1 s and then returned to the magnet. After another 10 minutes the process was repeated once more. An ¹³C-NMR spectrum of the solid sample was recorded and the signal was found to be in accordance with thermal equilibrium at 6.56 T and 2.5 K.

T₁-values for the ¹³C atom in 2-¹³C-2,2-bis(trideuteromethyl)-1,1,3,3-tetradeuteropropane-1,3-diol

H ₂ O, air saturated, 37°C, 7 T	133 s
H ₂ O, degassed, 37°C, 7 T	157 s
Human bloodplasma, 37°C, 7 T	96 s
Solid, 20°C, 7 T	237 s

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Solid, 2.5 K, 7 T

45 hours

Examples 8-9

Solution experiments

Example 8

A sample of solid 1-¹³C-2,2,2',2',2'',2'''-hexadeuterotris(hydroxymethyl)nitromethane (20 mg) was subjected to the above-mentioned pumping procedure (see Examples 5-7) and then in less than 1 second moved to a holding field of 0.4 T where also a sample of deuterium oxide (3 ml) at a temperature of 40°C, stirred by nitrogen bubbling, was kept. The solid was added to the liquid and a clear solution was obtained in less than 1 s. This solution was pipetted over to a 5 mm standard NMR-sample tube and moved to a nearby NMR-spectrometer while kept in a holding field of 10 mT. The sample was inserted into the spectrometer and a ¹³C-spectrum was recorded. The whole process of moving the sample out of the cryomagnet, dissolution, sample preparation, transport and spectroscopy took 35 s. The intensity of the ¹³C-signal was compared to the intensity after the sample had reached thermal equilibrium at 40°C and 7 T. An enhancement factor of 12 was found.

Example 9

A sample of solid 2-¹³C-2,2-bis(trideuteromethyl)-1,1,3,3-tetradeuteropropane-1,3-diol (20 mg) was subjected to the above-mentioned pumping procedure (see Examples 5-7) and then in less than 1 second moved to a holding field of 0.4 T where also a sample of deuterium oxide (3 ml) at a temperature of 40°C, stirred by nitrogen bubbling, was kept. The solid was added to the liquid and a clear solution was obtained in less than 1

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s. This solution was pipetted over to a 5 mm standard NMR-sample tube and moved to a nearby NMR-spectrometer while kept in a holding field of 10 mT. The sample was inserted into the spectrometer and a ^{13}C -spectrum was recorded. The whole process of moving the sample out of the cryomagnet, dissolution, sample preparation, transport and spectroscopy took 35 s. The intensity of the ^{13}C -signal was compared to the intensity after the sample had reached thermal equilibrium at 40°C and 7 T. An enhancement factor of 21 was found.

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Claims

1. A method of magnetic resonance investigation of a sample, preferably of a human or non-human animal body, said method comprising:

(i) producing a hyperpolarised solution of a high T_1 agent by dissolving in a physiologically tolerable solvent a hyperpolarised solid sample of said high T_1 agent;

(ii) where the hyperpolarisation of the solid sample of said high T_1 agent in step (i) is effected by means of a polarising agent, optionally separating the whole, substantially the whole, or a portion of said polarising agent from said high T_1 agent;

(iii) administering said hyperpolarised solution to said sample;

(iv) exposing said sample to a second radiation of a frequency selected to excite nuclear spin transitions in the MR imaging nuclei of the high T_1 agent;

(v) detecting magnetic resonance signals from said sample; and

(vi) optionally, generating an image, dynamic flow data, diffusion data, perfusion data, physiological data or metabolic data from said detected signals,

wherein said high T_1 agent in said hyperpolarised solution has a T_1 value (at a field strength in the range 0.01-5T and a temperature in the range 20-40°C) of at least 5 seconds.

2. A method as claimed in claim 1 wherein the polarisation of the solid high T_1 agent is retained during transportation in a magnetic field and low temperature.

3. A method as claimed in claim 2 wherein the magnetic

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field during said transportation is greater than 10 mT.

4. A method as claimed in claim 2 wherein the magnetic field during said transportation is greater than 0.1T.

5. A method as claimed in claim 2 wherein the magnetic field during said transportation is greater than 0.5T.

6. A method as claimed in claim 2 wherein the magnetic field during said transportation is greater than 1T.

7. A method as claimed in any one of claims 2 to 6 wherein the temperature during said transportation is lower than 80K.

8. A method as claimed in any one of claims 2 to 6 wherein the temperature during said transportation is lower than 4.2K.

9. A method as claimed in any one of claims 2 to 6 wherein the temperature during said transportation is lower than 1K.

10. A method as claimed in any one of the preceding claims wherein the hyperpolarised solution thus formed retains its polarisation during transportation in a magnetic field.

11. A method as claimed in claim 10 wherein the magnetic field during said transportation is greater than 10 mT.

12. A method as claimed in claim 10 wherein the magnetic field during said transportation is greater than 0.1T.

13. A method as claimed in claim 10 wherein the

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magnetic field during said transportation is greater than 0.5T.

14. A method as claimed in claim 10 wherein the magnetic field during said transportation is greater than 1T.

15. A method as claimed in any one of the preceding claims wherein said high T₁ agent is soluble in a physiologically tolerable carrier to a concentration of at least 1mM at a rate of 1mM/3T₁.

16. A method as claimed in claim 15 wherein said high T₁ agent is soluble in a physiologically tolerable carrier to a concentration of at least 1mM at a rate of 1mM/2T₁.

17. A method as claimed in claim 15 wherein said high T₁ agent is soluble in a physiologically tolerable carrier to a concentration of at least 1mM at a rate of 1mM/T₁.

18. A method as claimed in any one of the preceding claims wherein a magnetic field is present during the dissolution stage.

19. A method as claimed in claim 18 wherein said magnetic field is greater than 10 mT.

20. A method as claimed in claim 18 wherein said magnetic field is greater than 0.1T.

21. A method as claimed in claim 18 wherein said magnetic field is greater than 0.5T.

22. A method as claimed in claim 18 wherein said magnetic field is greater than 1T.

23. A method as claimed in any one of the preceding

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claims wherein step (i) comprises polarising a solid high T_1 agent by irradiating a polarising agent whereby to cause dynamic nuclear polarisation.

24. A method as claimed in claim 1 wherein step i) comprises polarising a hyperpolarisable gas before, during or after introducing a high T_1 agent thereto whereby to cause nuclear polarisation of said high T_1 agent.

25. A method as claimed in any one of the preceding claims wherein said high T_1 agent has a T_1 value (at a field strength of 0.01-5T and a temperature in the range 20-40°C) of at least 10 secs.

26. A method as claimed in any one of the preceding claims wherein said high T_1 agent has a T_1 value (at a field strength of 0.01-5T and a temperature in the range 20-40°C) of at least 30 secs.

27. A method as claimed in any one of the preceding claims wherein said high T_1 agent has a T_1 value (at a field strength of 0.01-5T and a temperature in the range 20-40°C) of at least 70 secs.

28. A method as claimed in any one of the preceding claims wherein said high T_1 agent has a T_1 value (at a field strength of 0.01-5T and a temperature in the range 20-40°C) of at least 100 secs.

29. A method as claimed in any one of the preceding claims wherein said high T_1 agent contains ^{13}C nuclei.

30. A method as claimed in any one of claim 1 to 28 wherein said high T_1 agent contains ^{15}N nuclei.

31. A method as claimed in any one of claim 1 to 28

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wherein said high T₁ agent contains ²⁹Si nuclei.

32. A method as claimed in any one of claim 1 to 28 wherein said high T₁ agent contains ³¹P nuclei.

33. A method as claimed in any one of claim 1 to 28 wherein said high T₁ agent contains ¹H nuclei.

34. A method as claimed in any one of claim 1 to 28 wherein said high T₁ agent contains ¹⁹F nuclei.

35. A method as claimed in any one of the preceding claims wherein said high T₁ agent exhibits a chemical shift of more than 2ppm per 2pH units or per Kelvin or upon being metabolised.

36. A method as claimed in any one of the preceding claims wherein said high T₁ agent exhibits a chemical shift of more than 10ppm per 2pH units or per Kelvin or upon being metabolised.

37. A method as claimed in claim 29 wherein said high T₁ agent has ¹³C at one particular position in its molecular structure in an amount above 5%.

38. A method as claimed in claim 29 wherein said high T₁ agent has ¹³C at one particular position in its molecular structure in an amount above 10%.

39. A method as claimed in claim 29 wherein said high T₁ agent has ¹³C at one particular position in its molecular structure in an amount above 25%.

40. A method as claimed in claim 29 wherein said high T₁ agent has ¹³C at one particular position in its molecular structure in an amount above 50%.

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41. A method as claimed in claim 29 wherein said high T₁ agent has ¹³C at one particular position in its molecular structure in an amount above 99%.

42. A method as claimed in claim 29 wherein said high T₁ agent is ¹³C enriched at one or more carbonyl or quaternary carbon positions.

43. A method as claimed in claim 42 wherein said high T₁ agent is deuterium labelled.

44. A method as claimed in claim 43 wherein said deuterium label is adjacent a ¹³C nucleus.

45. A method as claimed in claim 30 wherein said high T₁ agent has ¹⁵N at one particular position in its molecular structure in an amount above 5%.

46. A method as claimed in claim 30 wherein said high T₁ agent has ¹⁵N at one particular position in its molecular structure in an amount above 10%.

47. A method as claimed in claim 30 wherein said high T₁ agent has ¹⁵N at one particular position in its molecular structure in an amount above 25%.

48. A method as claimed in claim 30 wherein said high T₁ agent has ¹⁵N at one particular position in its molecular structure in an amount above 50%.

49. A method as claimed in claim 30 wherein said high T₁ agent has ¹⁵N at one particular position in its molecular structure in an amount above 99%.

50. A method as claimed in claim 31 wherein said high T₁ agent has ²⁹Si at one particular position in its molecular structure in an amount above 5%.

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51. A method as claimed in claim 31 wherein said high T₁ agent has ²⁹Si at one particular position in its molecular structure in an amount above 10%.
52. A method as claimed in claim 31 wherein said high T₁ agent has ²⁹Si at one particular position in its molecular structure in an amount above 25%.
53. A method as claimed in claim 31 wherein said high T₁ agent has ²⁹Si at one particular position in its molecular structure in an amount above 50%.
54. A method as claimed in claim 31 wherein said high T₁ agent has ²⁹Si at one particular position in its molecular structure in an amount above 99%.
55. A method as claimed in claim 24 wherein said hyperpolarisable gas is a noble gas.
56. A method as claimed in claim 55 wherein said noble gas is selected from the group consisting of He and Xe.
57. A method as claimed in claim 55 wherein said noble gas is ³He.
58. A method as claimed in claim 24 wherein said hyperpolarisable gas is used in liquid or solid form.
59. A method as claimed in claim 24 wherein said hyperpolarisable gas is a mixture of two or more gases.
60. A composition comprising a polarised ¹³C, ¹⁵N, ¹⁹F, ²⁹Si, ³¹P or ¹H enriched compound together with one or more physiologically acceptable carriers or excipients for use in a method as claimed in any one of the preceding claims.

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61. A contrast medium comprising a polarised high T_1 agent being enriched with ^{13}C , ^{15}N , ^{19}F , ^{29}Si , ^{31}P or ^1H nuclei having a T_1 of 2 secs or more in solution at magnetic fields of 0.01-10T, together with one or more physiologically acceptable carriers or excipients.

62. A contrast medium as claimed in claim 61 comprising a polarised high T_1 agent being enriched with ^{13}C , ^{15}N , ^{19}F , ^{29}Si , ^{31}P or ^1H nuclei having a T_1 of 10 secs or more in solution at magnetic fields of 0.01-10T, together with one or more physiologically acceptable carriers or excipients.

63. A contrast medium as claimed in claim 61 comprising a polarised high T_1 agent being enriched with ^{13}C , ^{15}N , ^{19}F , ^{29}Si , ^{31}P or ^1H nuclei having a T_1 of 30 secs or more in solution at magnetic fields of 0.01-10T, together with one or more physiologically acceptable carriers or excipients.

64. A contrast medium as claimed in claim 61 comprising a polarised high T_1 agent being enriched with ^{13}C , ^{15}N , ^{19}F , ^{29}Si , ^{31}P or ^1H nuclei having a T_1 of 60 secs or more in solution at magnetic fields of 0.01-10T, together with one or more physiologically acceptable carriers or excipients.

65. A method as claimed in claim 1 wherein step i) comprises:

- i) subjecting said high T_1 agent to a high magnetic field at low temperature;
- ii) exposing the agent to a T_1 shortening effect in order to attain thermodynamic equilibrium at said low temperature.

66. A method as claimed in claim 65 wherein said magnetic field is greater than 1T.

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67. A method as claimed in claim 65 wherein said magnetic field is 5T or more.

68. A method as claimed in claim 65 wherein said magnetic field is 15T or more.

69. A method as claimed in any one of claims 65 to 68 wherein said low temperature is 100K or less.

70. A method as claimed in any one of claims 65 to 68 wherein said low temperature is 4.2K or less.

71. A method as claimed in any one of claims 65 to 68 wherein said low temperature is 1K or less.

72. A method as claimed in any one of claims 65 to 68 wherein said low temperature is 0.1K or less.

73. A method as claimed in any one of claims 65 to 72 wherein said T_1 shortening effect is provided by exposure to a variable magnetic field gradient.

74. A method as claimed in any one of claims 65 to 72 wherein said T_1 shortening effect is provided by field cycling to a field allowing cross polarisation.

75. A method as claimed in any one of claims 65 to 72 wherein said T_1 shortening effect is provided by gradually increasing the magnetic field at such a rate that the increase in polarisation of the high T_1 agent is maximised.

76. A method as claimed in any one of claims 65 to 72 wherein said T_1 shortening effect is provided by gradually decreasing the temperature at such a rate that the increase in polarisation of the high T_1 agent is maximised.

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77. A method as claimed in any one of claims 65 to 76 wherein said T₁ shortening effect is provided by adding a material with unpaired electrons during the period when the agent is exposed to low temperature.

78. A method as claimed in claim 77 wherein said magnetic material is selected from the group consisting of O₂ or NO.

79. An administrable composition comprising a polarised high T₁ agent prepared as described in any one of claims 65 to 78 dissolved in an aqueous solvent.

80. A composition as claimed in claim 79 wherein said aqueous solvent is also polarised.

81. A composition as claimed in either one of claims 79 and 80 wherein said aqueous solvent is water.

82. A composition as claimed in any one of claims 79 to 81 comprising a polarised ¹³C, ¹⁵N, ¹⁹F, ²⁹Si, ³¹P or ¹H enriched compound together with one or more physiologically acceptable carriers or excipients.

83. A contrast medium comprising a polarised high T₁ agent prepared as described in any one of claims 65 to 78 enriched with ¹³C nuclei having a T₁ relaxation time of 2s or more in solution at magnetic fields of 0.005-10T, together with one or more physiologically acceptable carriers or excipients.

84. A method of magnetic resonance investigation of a sample, preferably of a human or non-human animal body, said method comprising:

i) producing solid hyperpolarised ¹²⁹Xe by irradiating a polarising agent whereby to cause dynamic nuclear polarisation.

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85. A method as claimed in claim 84 wherein said polarising agent is a substance containing an unpaired electron.

86. A method as claimed in claim 85 wherein said polarising agent comprises nitroxide, trityl or Cr(V) moieties.

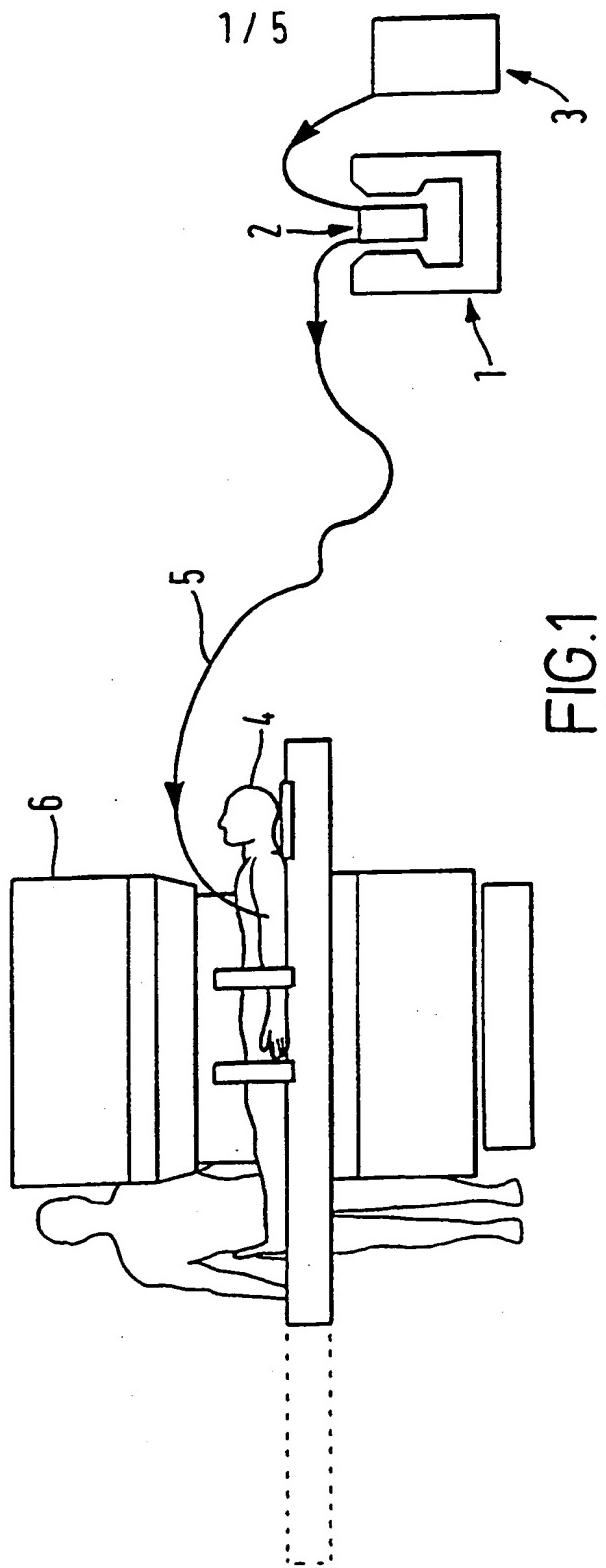


FIG.1

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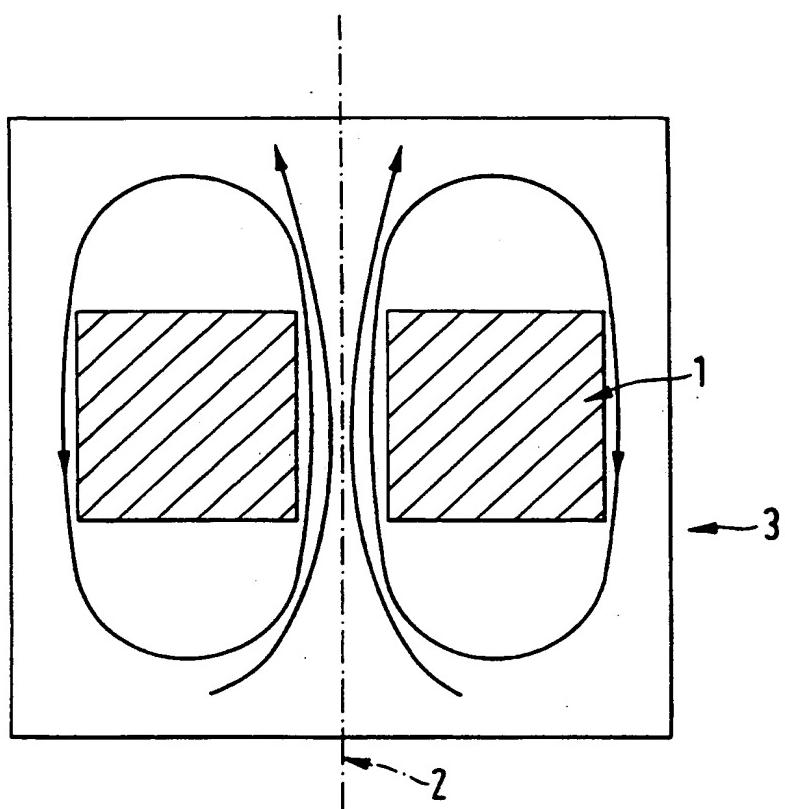


FIG.2

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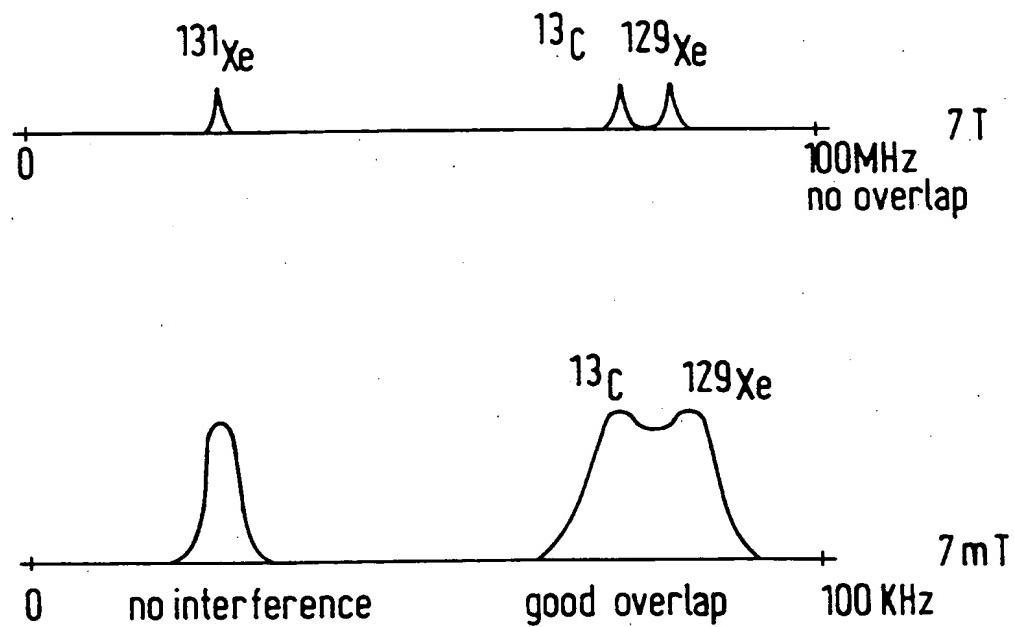


FIG.3

BUILD UP OF MAGNETIZATION VERSUS TIME

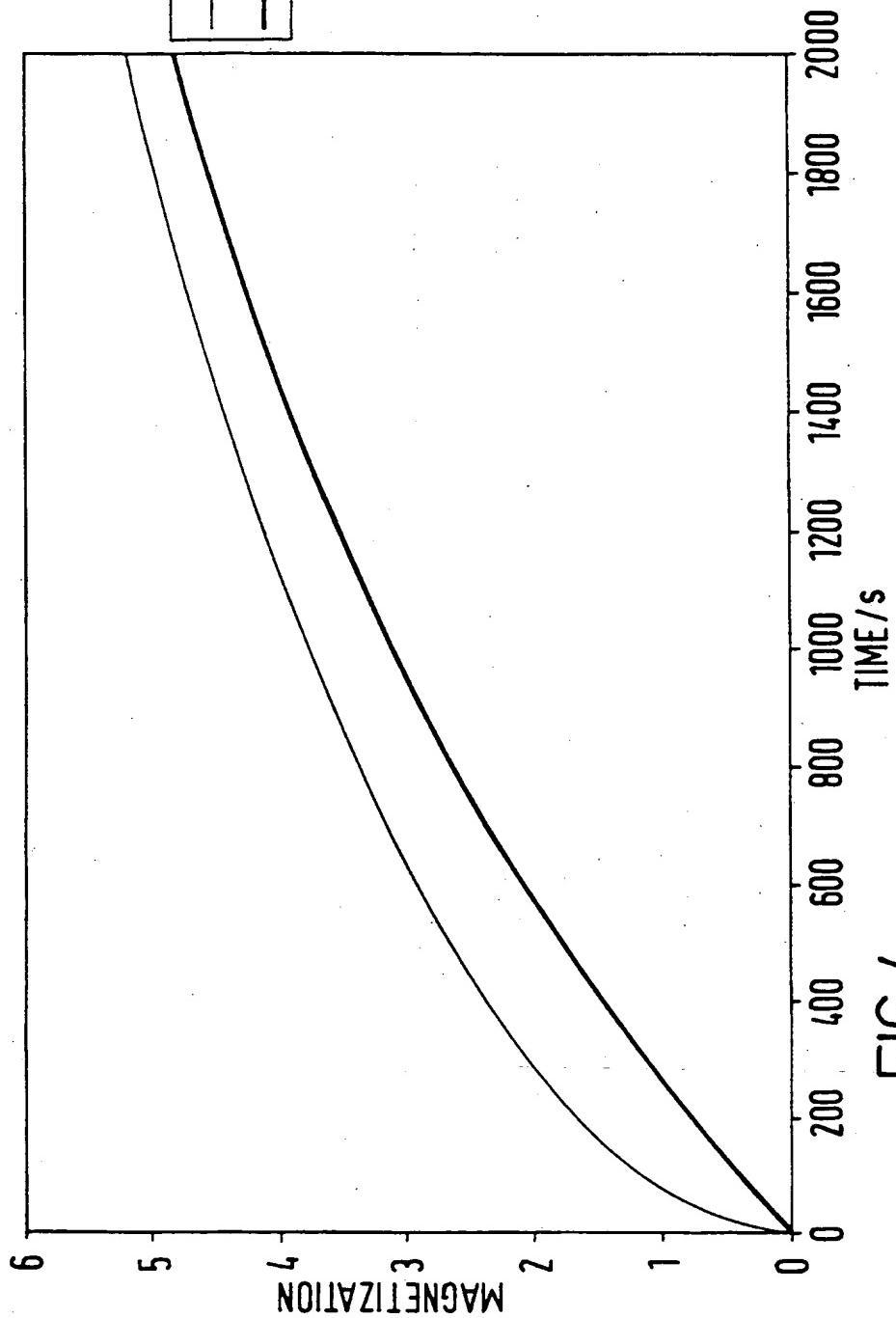


FIG. 4

— FIELD

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MAGNETIC FIELD VERSUS TIME

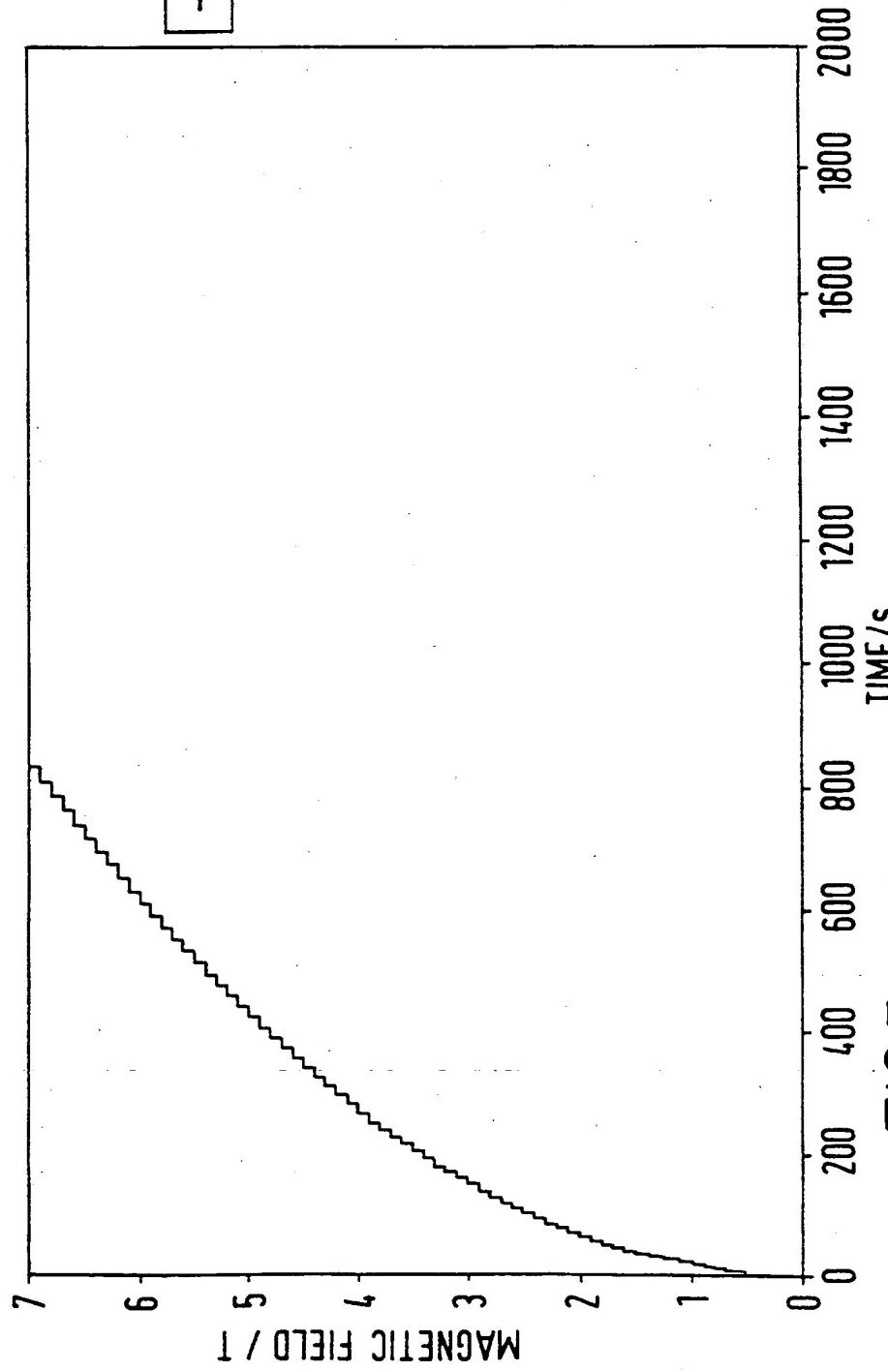


FIG.5

INTERNATIONAL SEARCH REPORT

Int'l. Application No
PCT/GB 98/03904

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 G01R33/28

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 G01R

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P, X	<p>WO 98 01766 A (NYCOMED) 15 January 1998</p> <p>see page 4 - page 5</p> <p>see page 10 - page 12</p> <p>---</p> <p>WO 97 37239 A (NAVON GIL ;BIFONE ANGELO (IT); ROOM TOOMAS (US); APPELT STEPHAN (U) 9 October 1997</p> <p>see page 2, line 36 - page 3, line 27</p> <p>see page 10, line 11 - line 36</p> <p>see page 11, line 17 - line 20</p> <p>---</p>	1,23-64, 84-86
X	<p>WO 97 37239 A (NAVON GIL ;BIFONE ANGELO (IT); ROOM TOOMAS (US); APPELT STEPHAN (U) 9 October 1997</p> <p>see page 2, line 36 - page 3, line 27</p> <p>see page 10, line 11 - line 36</p> <p>see page 11, line 17 - line 20</p> <p>---</p>	1,2
P, X	<p>WO 98 30918 A (COCKBAIN JULIAN R M ;NYCOMED IMAGING AS (NO); PETTERSSON STEFAN (S) 16 July 1998</p> <p>see page 1 - page 9</p> <p>---</p> <p>-/-</p>	1,61-64

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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Date of the actual completion of the international search

6 April 1999

Date of mailing of the international search report

27/04/1999

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Diot, P

INTERNATIONAL SEARCH REPORT

International Application No
PCT/GB 98/03904

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	US 5 617 859 A (CLINE HARVEY E ET AL) 8 April 1997 cited in the application see column 5 ---	65
A	PIETRASS T ET AL: "Optically polarized /sup 129/Xe in NMR spectroscopy" ADVANCED MATERIALS, OCT. 1995, VCH VERLAGSGESELLSCHAFT, GERMANY, vol. 7, no. 10, pages 826-838, XP002098966 ISSN 0935-9648 Chapter: 6. Polarization transfer ---	84

INTERNATIONAL SEARCH REPORT

Information on patent family members

Int'l. Application No
PCT/GB 98/03904

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